

Mouse Urocortin 2 ELISA Kit

Catalog Number: YK190 (1 x 96 wells)

For Research Use Only. Not for use in diagnostic procedures.

v. 1.0

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1. Introduction

Urocortin 2 (Ucn 2), also known as stresscopin-related peptide, is a novel predicted neuropeptide related to corticotropin-releasing factor (CRF). The peptide consisting of 38 amino acid residues was first demonstrated to be expressed centrally and to bind selectively to type 2 CRF receptor (CRFR2)¹⁾. In the rodent, Ucn 2 transcripts were shown to be expressed in the discrete regions of the central nervous system including stress-related cell groups in the hypothalamus and brainstem¹⁾. More recently, the expression of Ucn 2 transcripts was detected in the olfactory bulb, pituitary, cortex, hypothalamus, and spinal cord²⁾. Ucn 2 mRNA was also found to be expressed widely in a variety of peripheral tissues, most highly in the skin and skeletal muscle tissues³⁾. Ucn 2-like immunoreactivity was detected by RIA in acid extracts of mouse brain, muscle, and skin³⁾. Immunohistochemically Ucn 2 was found in both skin epidermis and adnexal structures and in the skeletal muscle myocytes³⁾. Ucn 2 gene transcription was stimulated in the hypothalamus and brainstem glucocorticoid administration to the mouse and inhibited by removal of glucocorticoids by adrenalectomy, suggesting a putative link between the CRFR1 and CRFR2 pathways²⁾. On the other hand, in the rat a stressor-specific regulation of Ucn 2 mRNA expression in the hypothalamic paraventricular nucleus was demonstrated, which raised the possibility of a modulary role of Ucn 2 mRNA in stress-induced alteration of anterior and posterior pituitary function, depending on the type of stress⁴⁾. Administration of dexamethasone to the mouse resulted in a decrease of Ucn 2 mRNA levels in the back skin region. Adrenalectomy significantly increased Ucn 2 mRNA levels in the skin, and the levels were reduced back to normal levels after corticoid replacement 3).

CRFR2 is found in cardiomyocytes and in endothelial and smooth muscle cells of the systemic vasculature. Ucn 2 is expressed in the mouse cardiomyocytes. In the mouse, Ucn 2 treatment augmented heart rate, exhibited potent inotropic and lusitropic actions on the left ventricle, and induced a downward shift of the diastolic pressure-volume relation⁵⁾. Ucn 2 also reduced systemic arterial pressure, associated with a lowering of systemic arterial elastance and systemic vascular resistance. The effects of Ucn 2 were specific to CRFR2 function and independent of beta-adrenergic receptors. These experiments demonstrated the potent cardiovascular physiologic actions of Ucn 2 in the both wild-type and cardiomyopathic mice and support a potential beneficial use of Ucn 2 in congestive heart failure treatment⁵⁾. The use of Ucn 2 was also proposed to treat ischemic heart disease because of its potent cardioprotective effect in the mouse heart and its minimal impact on the hypothalamic stress axis⁶⁾.

Administration of Ucn 2 to the mouse prevented the loss of skeletal muscle mass resulting from disuse due to casting, corticosteroid treatment, and nerve damage. In addition, Ucn 2 treatment prevented the loss of skeletal muscle force and myocyte cross-sectional area that accompanied muscle mass losses resulting from disuse due to casting. In normal muscles of the mouse, Ucn 2 increased skeletal muscle mass and force. It was thus proposed that Ucn 2 might find utility



in the treatment of skeletal muscle wasting diseases including age-related muscle loss or sarcopenia⁷⁾.

Mouse urocortin 2 (Ucn 2) is a new peptide predicted from mouse cDNA sequence and its physiologic and pathophysiologic significance has not yet been fully elucidated. However, the experimental data presented to date provided evidence for the important physiologic roles of Ucn 2 and urge the necessity of further investigation of the peptide from various points of view.

We succeeded this time in the development of mouse urocortin 2 EIA kit which highly specific for mouse Ucn 2 with almost no crossreaction to Ucn 1 (mouse, rat), Ucn 3 (mouse), ACTH (mouse, rat) and CRF (mouse, rat, human). The kit can be used for measurement of Ucn 2 in mouse plasma or serum with high sensitivity. It will be a specifically useful tool for Ucn 2 research.

YK190 Mouse Urocortin 2 EIA Kit

- ▼ The assay kit can measure mouse urocortin 2 within the range of 0.82-200 ng/mL.
- ▼ The assay is completed within 16-18 hr. +3 hr.
- ▼ With one assay kit, 41 samples can be measured in duplicate
- Test sample: Mouse plasma & serum Sample volume: 20 μL
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- The 96-well plate of this kit consists of 12 8-wells strips, so that divided use by the strips is possible at user's option.
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Precision and reproducibility

Intra-assay CV (%)

Mouse plasma 2.51-5.25 Mouse serum 6.71-9.01

Inter-assay CV (%)

Mouse plasma 4.70-8.28 Mouse serum 6.36-11.12

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Inter-assay CV (%)

Mouse plasma 4.70-8.28 Mouse serum 6.36-11.12

Stability and Storage

Store all of the components at 2-8°C.

The kit is stable under the condition for 18 months from the date of manufacturing.

The expiry date is stated on the package.

Contents

- 1) Antibody coated plate
- 2) Standard
- 3) Labeled antigen
- 4) SA-HRP solution
- 5) Substrate buffer
- 6) OPD tablet
- 7) Stopping solution
- 8) Buffer solution
- 9) Washing solution (concentrated)
- 10 Adhesive foil)

2. Characteristics

This Eagle Biosciences Mouse Urocortin 2 ELISA Assay kit is used for quantitative determination of urocortin 2 in mouse plasma & serum samples. The kit is characterized by its sensitive quantification and high specificity. In addition, it has no influence by other components in samples. Mouse urocortin 2 standard is highly purified synthetic product. This Eagle Biosciences Mouse Urocortin 2 ELISA Assay Kit is for research use only and not to be used for therapeutic procedures.

Specificity

This Eagle Biosciences Mouse Urocortin 2 ELISA Assay kit has high specificity to mouse urocortin 2 and shows cross reactivity neither urocortin 1 (mouse, rat), urocortin 3 (mouse), ACTH (mouse, rat) nor CRF (mouse, rat, human).

Assay principle

This Eagle Biosciences Mouse GLP-2 ELISA Assay Kit for the determination of mouse urocortin 2 in samples is based on a competitive enzyme immunoassay using combination of highly specific antibody to mouse urocortin 2 and biotin-avidin affinity system. To the wells of plate coated with rabbit anti mouse urocortin 2 antibody, standard or samples, labeled antigen are added for competitive immunoreaction. After incubation and plate washing, horse radish peroxidase (HRP) labeled streptoavidin (SA) is added to form HRP labeled streptoavidin-biotinylated antigen-antibody complex on the surface of the wells. Finally, HRP enzyme activity is determined by o-Phenylenediamine dihydrochloride (OPD) and the concentration of mouse urocortin 2 is calculated.

3. Composition

Component	Form	Quantity	Main Ingredient
Antibody coated plate	Microtiter plate	1 plate (96 wells)	Rabbit anti mouse urocortin 2 antibody
Standard	Lyophilized	1 vial (200ng/vial)	Synthetic mouse urocortin2
Labeled antigen	Lyophilized	1 vial	Biotinylated mouse urocortin 2
SA-HRP solution	Liquid	1 bottle (12 mL)	HRP labeled streptavidin
Substrate buffer	Liquid	1 bottle (24 mL)	0.015% hydrogen peroxide
OPD tablet	Tablet	2 tablets	o-Phenylenediamine dihydrochloride
Stopping solution	Liquid	1 bottle (12 mL)	$1M^{3}H_{2}SO_{4}$
Buffer solution	Liquid	1 bottle (15 mL)	Phosphate buffer
Washing solution (concentrated)	Liquid	1 bottle (50 mL)	Concentrated saline
Adhesive foil		3 pieces	

4. Method

Equipment required

- 1. Photometer for microtiter plate (plate reader), which can read extinction 2.5 at 492 nm
- 2. Microtiter plate shaker
- 3. Washing device for microtiter plate and dispenser with aspiration system
- 4. Micropipettes, multi-channel pipettes for 8 wells or 12 wells and their tips
- 5. Test tubes for preparation of standard solution
- 6. Graduated cylinder (1,000 mL)
- 7. Distilled or deionized water

5. Preparatory work

1. Preparation of standard solution:

Reconstitute the mouse urocortin 2 standard with 1 mL of buffer solution, which affords 200ng/mL standard solution. The reconstituted standard solution (0.1mL) is diluted with 0.2 mL of buffer solution that yields 66.7ng/mL standard solution. Repeat the dilution procedure to make each standard solution of 22.2, 7.41, 2.47, 0.82 ng/mL. Buffer solution itself is used as 0ng/mL.

2. Preparation of labeled antigen:

Reconstitute labeled antigen with 6 mL of distilled water.

3. Preparation of substrate solution:

Resolve one OPD tablet with 11 mL of substrate buffer. It should be prepared immediately before use.

4. Preparation of washing solution:

Dilute 50 mL of washing solution (concentrated) to 1000 mL with distilled or deionized water.

5. Other reagents are ready for use.

6. Procedure

- 1. Before start assay, bring all the reagents and samples to room temperature $(20 \sim 30^{\circ}\text{C})$.
- 2. Add 0.35mL/well of washing solution into the wells and aspirate the washing solution in the wells. Repeat this washing procedure further twice (total 3 times).

- Finally, invert the plate and tap it onto an absorbent surface, such as paper toweling, to ensure blotting free of most residual washing solution.
- 3. Fill 25μ L of buffer solution into the wells first, then introduce 20μ L of each of standard solutions (0, 0.82, 2.47, 7.41, 22.2, 66.7, 200 ng/mL) or samples and finally add 50μ L of labeled antigen into the wells. The total pipetting time of standard solutions and samples for a whole plate should not exceed 30 min.
- 4. Cover the plate with adhesive foil and incubate it at 4° C overnight for $16 \sim 18$ hours. (Still, plate shaker not need)
- 5. After incubation, move the plate back to room temperature keeping for about 40 minutes and take off the adhesive foil, aspirate and wash the wells four times with approximately 0.35 mL/well of washing solution. Finally, invert the plate and tap it onto an absorbent surface, such as paper toweling, to ensure blotting free of most residual washing solution.
- 6. Pipette 100μL of SA-HRP solution into each of the wells.
- 7. Cover the plate with adhesive foil and incubate it at room temperature (20 ~ 30°C) for 2 hours. During the incubation, the plate should be shake with a plate shaker.
- 8. Resolve OPD tablet with 11 mL of substrate buffer. It should be prepared immediately before use.
- 9. Take off the adhesive foil, aspirate and wash the wells four times with approximately 0.35 mL/well of washing solution. Finally, invert the plate and tap it onto an absorbent surface, such as paper toweling, to ensure blotting free of most residual washing solution.
- 10. Add $100\mu L$ of substrate solution to each of the wells, cover the plate with adhesive foil and incubate it for 20 minutes at room temperature.
- 11. Add 100μL of stopping solution into each of the wells to stop color reaction.
- 12. Read the optical absorbance of the solution in the wells at 492 nm. Calculate mean absorbance values of wells containing standards and plot a standard curve on semilogarithmic graph paper (abscissa: concentration of standard; ordinate: absorbance values). Use the average absorbance of each sample to determine the corresponding value by simple interpolation from this standard curve

7. Notes

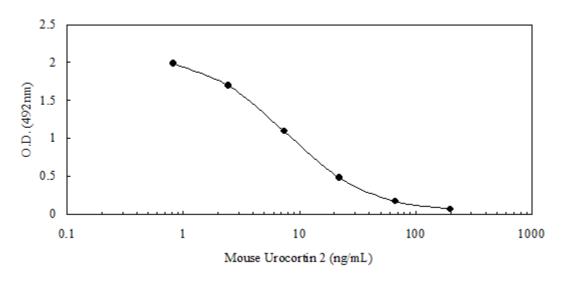
1. EDTA-2Na additive blood collection tube is recommended for the plasma collection. Serum and plasma samples must be used as soon as possible after collection. If the

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- samples are tested later, they should be divided into test tubes in small amount and frozen at or below -30°C. Avoid repeated freezing and thawing of samples.
- 2. Standard and labeled antigen solutions should be prepared immediately before use. This kit can be use dividedly in strips. In such case, the rest of reconstituted reagents (standard and labeled antigen) should be stored at -30°C.
- 3. The total pipetting time of standard solutions and samples for a whole plate should not exceed 30 min.
- 4. During storage of washing solution (concentrated) at 2-8°C, precipitates may be observed, however they will be dissolved when diluted. Diluted washing solution is stable for 6 months at 2-8°C.
- 5. Pipetting operations may affect the precision of the assay, pipette standard solutions or samples precisely into each well of plate. In addition, using clean test tubes or vessels in assay and use new tip for each standard or sample to avoid cross contamination.
- 6. When sample value exceeds 200 ng/mL, it needs to be diluted with buffer solution to proper concentration.
- 7. During incubation in the room temperature except color reaction, the test plate should be shake gently by plate shaker to promote immunoreaction.
- 8. Perform all the determination in duplicate.
- 9. Read plate optical absorbance of reaction solution in wells as soon as possible after stopping color reaction.
- 10. To quantitate accurately, always run a standard curve when testing samples.
- 11. Protect reagents from strong light (e.g. direct sunlight) during storage and assay.
- 12. Satisfactory performance of the test is guaranteed only when reagents are used from combination pack with identical lot number.

8. Performance Characteristics

Typical Standard Curve



Analytical recovery

Mouse Plasma A

Added Mouse Urocortin 2	Observed	Expected	Recovery
(ng/mL)	(ng/mL)	(ng/ml)	(%)
0.0	1.58		
1.0	2.92	2.58	113.18
5.0	7.36	6.58	111.85
30.0	35.82	31.58	113.43
50.0	59.92	51.58	116.17

Mouse Plasma B

Added Mouse Urocortin 2	Observed	Expected	Recovery
(ng/mL)	(ng/mL)	(ng/ml)	(%)
0.0	1.72		
1.0	2.71	2.72	99.63
5.0	6.73	6.72	100.15
30.0	35.99	31.72	113.46
50.0	60.79	51.72	117.54

Mouse Plasma C

Added Mouse Urocortin 2	Observed	Expected	Recovery
(ng/mL)	(ng/mL)	(ng/ml)	(%)
0.0	1.67		
1.0	2.64	2.67	98.88
5.0	7.07	6.67	106.00
30.0	30.89	31.67	97.54
50.0	55.80	51.67	107.99
Mouse Plasma D			
Added Mouse Urocortin 2	Observed	Expected	Recovery
(ng/mL)	(ng/mL)	(ng/ml)	(%)
0.0	1.30		
1.0	2.62	2.30	113.91
5.0	7.11	6.30	112.86
30.0	32.96	31.30	105.30
50.0	49.97	51.30	97.41
Mouse Serum A			
Added Mouse Urocortin 2	Observed	Expected	Recovery
(ng/mL)	(ng/mL)	(ng/ml)	(%)
0.0	2.69		
1.0	4.02	3.69	108.94
5.0	8.57	7.69	111.44
30.0	38.24	32.69	116.98
50.0	70.07	52.69	132.99
Mouse Serum B			
Added Mouse Urocortin 2	Observed	Expected	Recovery
(ng/mL)	(ng/mL)	(ng/ml)	(%)
0.0	2.66		
1.0	3.91	3.66	106.83
5.0	8.78	7.66	114.62
30.0	44.14	32.66	135.15
50.0	78.51	52.66	149.09

Mouse Serum C

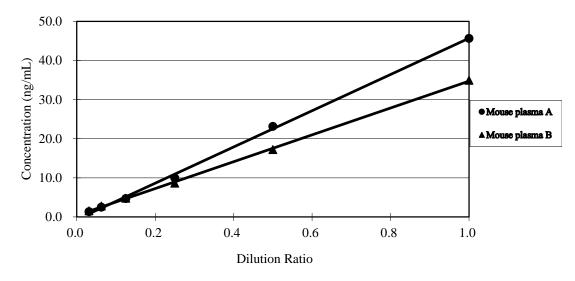
Added Mouse Urocortin 2	Observed	Expected	Recovery
(ng/mL)	(ng/mL)	(ng/ml)	(%)
0.0	2.96		
1.0	4.14	3.96	104.55
5.0	9.12	7.96	114.57
30.0	43.45	32.96	131.83
50.0	78.94	52.96	149.06

Mouse Serum D

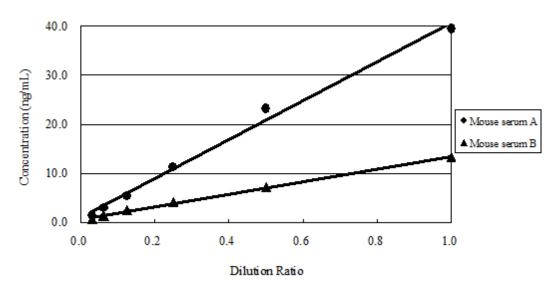
Added Mouse Urocortin 2	Observed	Expected	Recovery
(ng/mL)	(ng/mL)	(ng/ml)	(%)
0.0	2.51		
1.0	3.59	3.51	102.28
5.0	8.48	7.51	112.92
30.0	38.72	32.51	119.10
50.0	71.82	52.51	136.77

Dilution Test

Mouse Plasma



Mouse Serum



Cross-Reactivity

Related peptides	Crossreactivity (%)
Urocortin 2 (mouse)	100
Urocortin 1 (mouse, rat)	0
Urocortin 3 (mouse)	0
ACTH (mouse, rat)	0.61
CRF (mouse, rat, human)	0

Precision and reproducibility

Test Sample	Intra-assay CV(%)	Inter-assay CV(%)
Mouse Plasma	2.51-5.25	4.70-8.28
Mouse Serum	6.71-9.01	6.36-11.12

Assay range: 0.82 ~ 200 ng/mL

9. Stability and Storage

Storage Store all of the components at 2-8°C.

Shelf life The kit is stable under the condition for 18 months from the date of

manufacturing. The expiry date is stated on the package

Package For 96 tests per one kit including standards

10. References

- Reyes TM. (2001) Urocortin II: a member of the corticotropin-releasing factor (CRF) neuropeptide family that is selectively bound by type 2 CRF receptors. *Proc Natl Acad Sci USA*. 98, 2843-2848
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- 4. Tanaka Y. (2003) Effect of stress and adrenalectomy on urocortin II mRNA expression in the hypothalamic paraventricular nucleus of the rat. *Neuroendocrinology.*, **78**, 1-11
- 5. Bale TL. (2004) The cardiovascular physiologic actions of urocortin II: acute effects in murine heart failure. *Proc Natl acad Sci U S A.,* **101,** 3697-3702
- 6. Brar BK. (2004) Urocortin II and urocortin III are cardioprotective against ischemia reperfusion injury: an essential endogenous cardioprotective role for corticotropin releasing factor receptor type 2 in the murine heart. *Endocrinology.*, **145**, 24-35
- 7. Hinkle RT. (2003) Urocortin II treatment reduces skeletal muscle mass and function loss during atrophy and increases nonatrophying skeletal muscle mass and function. *Endocrinology.*, **144**, 4939-4946

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For further information about this kit, its application or the procedures in this kit, please contact the Technical Service Team at Eagle Biosciences, Inc. at info@eaglebio.com or at 866-411-8023.

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