Human Anti-Mouse Antibody (HAMA) ELISA

Catalog Number:
HAM31-K01 (1 x 96 wells)
For Research Use Only. Not for use in diagnostic procedures.
v. 2.0 (07.26.15)
INTENDED USE
The Eagle Biosciences Human Anti-Mouse Antibody (HAMA) ELISA Assay Kit (enzyme-linked immunosorbent kit) is intended for the quantitative determination of human Anti-Mouse Antibody (HAMA) levels in serum. The Eagle Biosciences Human Anti-Mouse Antibody (HAMA) ELISA Assay Kit is for research use only and not to be used in diagnostic procedures.

INTRODUCTION
Clinically, mouse monoclonal antibodies (IgG) and their fragments are used in vivo diagnosis procedure (radionuclides) and treatment for patients with various diseases. In patients, even a single dose injection of murine monoclonal IgG may induce immune response directed against this foreign protein (immunogen). In the circulation, the presence of human antibody against murine IgG would bind to the injected murine IgG and, therefore, diminish the efficacy of either in-vivo diagnosis or treatment. Especially, the HAMA would increase the risk of anaphylactic complications to subsequent administration of the murine IgG based therapy.

The present of HAMA in patient serum or plasma specimens causes both false positive and false negative immunoassay test results depending on assay principles and monoclonal antibodies use in the assay system.

This HAMA ELISA is a ready to use test kit with well-breakable microtiter plate and simple test procedures. It also provides a wide measurement range without high dose “hook” effect.

PRINCIPLE OF THE ASSAY
The Eagle Biosciences Human Anti-Mouse Antibody (HAMA) ELISA Assay Kit is designed, developed and produced for the quantitative measurement of HAMA in serum and plasma samples. The assay utilizes the two-site “sandwich” technique with two selected antibodies that bind to HAMA.

Assay standards, controls and patient samples are directly added to wells of a microplate that is coated with murine IgG. After the first incubation period, the HAMA binds to the murine IgG on the wall of microtiter well and unbound proteins in each microtiter well are washed away. Then a horseradish peroxidase (HRP) labeled murine IgG is added to each microtiter well and a “sandwich” of “murine IgG HAMA – murine IgG” is formed. The unbound HRP conjugated murine IgG is removed in the subsequent washing step. For the detection of this immunocomplex, the well is then incubated with a substrate solution in a timed reaction and then measured in a spectrophotometric microplate reader. The enzymatic activity of the immunocomplex bound to HAMA on the wall of the microtiter well is directly proportional to the amount of HAMA in the sample. A standard curve is generated by plotting the absorbance versus the respective HAMA concentration for each standard on point-to-point, cubical scales or 4 parameter curve fit. The concentration of HAMA in test samples is determined directly from this standard curve.
REAGENTS: Preparation and Storage
This test kit must be stored at 2 – 8°C upon receipt. For the expiration date of the kit refer to the label on the kit box. All components are stable until the expiration date.

Allow all reagents to come to room temperature prior to use. Regents from different kit lot numbers should not be combined or interchanged.

1. **Murine IgG Coated Microplate**
   One well breakable microplate with 12 x eight strips (96 wells total) coated with murine IgG. The plate is framed and sealed in a foil zipper bag with a desiccant. This reagent should be stored at 2 – 8°C and is stable until the expiration date on the kit box.

2. **HAMA Tracer Antibody**
   One vial containing 0.6 mL HRP labeled murine IgG in a stabilized protein matrix. This reagent must be diluted with Tracer Antibody Diluent before use. This reagent should be stored at 2 – 8°C and is stable until the expiration date on the kit box.

3. **Tracer Antibody Diluent**
   One vial containing 12 mL ready to use buffer. This should be used only for tracer antibody dilution according to the assay procedures. This reagent should be stored at 2 – 8°C and is stable until the expiration date on the kit box.

4. **Assay Buffer**
   One bottle containing 30 mL of ready to use phosphate buffered saline based assay buffer with bovine serum albumin added. This reagent should be stored at 2 – 8°C and is stable until the expiration date on the kit box.

5. **ELISA Wash Concentrate**
   One bottle contains 30 mL of 30 fold concentrate. Before use the contents must be diluted with 870 mL of distilled water and mix well. Upon dilution this yields a working wash solution containing a surfactant in phosphate buffered saline with a non-azide and non-mercury based preservative. The diluted wash buffer should be stored at room temperature and is stable until the expiration date on the kit box.

6. **ELISA HRP Substrate**
   One bottle contains 12 mL of tetramethylbenzidine (TMB) with stabilized hydrogen peroxide. This reagent should be stored at 2 – 8°C and is stable until the expiration date on the kit box.

7. **ELISA Stop Solution**
   One bottle contains 12 mL of 0.5 M sulfuric acid. This reagent should be stored at 2 – 8°C or room temperature and is stable until the expiration date on the kit box.

8. **HAMA Standards**
   Five vials containing different levels of HAMA in a liquid protein matrix with a non-azide based preservative. Refer to vial for exact concentration for each standard. These reagents should be stored at 2 – 8°C and are stable until the expiration date on the kit box.

Human Anti-Mouse Antibody (HAMA) ELISA Assay Kit
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9. **HAMA Controls**

Two vials containing different levels of HAMA in a liquid protein matrix with a non-azide based preservative. **Refer to vials for exact concentration range for each control.** Both controls should be stored at 2 – 8°C and are stable until the expiration date on the kit box.

**SAFETY PRECAUTIONS**

The Human Anti-Mouse Antibody (HAMA) Assay Kit reagents must be used in a professional laboratory environment and is for Research Use Only and is not to be used in diagnostic procedures. Only source material from which reagents of bovine serum was derived in the contiguous 48 United States. It was obtained only from donor health animals maintained under veterinary supervision and found free of contagious diseases. Wear gloves while performing this assay and handle these reagents as if they were potentially infectious. Avoid contact with reagents containing TMB, hydrogen peroxide, or sulfuric acid. TMB may cause irritation to skin and mucous membranes and cause an allergic skin reaction. TMB is a suspected carcinogen. Sulfuric acid may cause severe irritation on contact with skin. Do not get in eyes, on skin, or on clothing. Do not ingest or inhale fumes. On contact, flush with copious amounts of water for at least 15 minutes. Use Good Laboratory Practices.

**MATERIALS REQUIRED BUT NOT PROVIDED**

- Precision single channel pipettes capable of delivering 25 µL, 50 µL, 100 µL, and 1000 µL
- Repeating dispenser suitable for delivering 100 µL.
- Disposable pipette tips suitable for above volume dispensing.
- Disposable 12 x 75 mm or 13 x 100 glass tubes.
- Disposable plastic 100 mL and 1000 mL bottle with caps.
- Aluminum foil.
- Deionized or distilled water.
- Plastic microtiter well cover or polyethylene film.
- ELISA multichannel wash bottle or automatic (semi-automatic) washing system.
- Spectrophotometric microplate reader capable of reading absorbance at 450 nm.

**SPECIMEN COLLECTION**

Only 50 µL of human serum or plasma is required for HAMA measurement in duplicate. No special preparation of individual is necessary prior to specimen collection. In the case of serum, whole blood should be collected and must be allowed to clot for a minimum of 30 minutes at room temperature before the serum is separated by centrifugation (850 – 1500xg for 10 minutes). The serum should be separated from the clot within three hours of blood collection and transferred to a clean test tube. Serum or plasma samples should be stored at 2 - 8°C if the assay is to be performed within 72 hours. Otherwise, patient samples should be stored at - 20°C or below until measurement. Avoid repeated (more than three times) freezing and thawing of specimen.

**Reagent Preparation**

1. Prior to use allow all reagents to come to room temperature. Reagents from different kit lot numbers should not be combined or interchanged.
2. ELISA Wash Concentrate must be diluted to working solution prior use. Please see REAGENTS section for details.
Assay Procedure

1. Place a sufficient number of murine IgG coated microwell strips/wells in a holder to run HAMA standards, controls and unknown samples in duplicate.

2. Test Configuration

<table>
<thead>
<tr>
<th>ROW</th>
<th>STRIP 1</th>
<th>STRIP 2</th>
<th>STRIP 3</th>
</tr>
</thead>
<tbody>
<tr>
<td>A</td>
<td>STD 1</td>
<td>STD 5</td>
<td>SAMPLE 2</td>
</tr>
<tr>
<td>B</td>
<td>STD 1</td>
<td>STD 5</td>
<td>SAMPLE 2</td>
</tr>
<tr>
<td>C</td>
<td>STD 2</td>
<td>C1</td>
<td>SAMPLE 3</td>
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<tr>
<td>D</td>
<td>STD 2</td>
<td>C1</td>
<td>SAMPLE 3</td>
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<td>E</td>
<td>STD 3</td>
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<tr>
<td>F</td>
<td>STD 3</td>
<td>C2</td>
<td>SAMPLE 4</td>
</tr>
<tr>
<td>G</td>
<td>STD 4</td>
<td></td>
<td>SAMPLE 1</td>
</tr>
<tr>
<td>H</td>
<td>STD 4</td>
<td></td>
<td>SAMPLE 1</td>
</tr>
</tbody>
</table>

3. Add 25 µL of standards, controls and patient samples into the designated microwell.
4. Add 100 µL of assay buffer to each well
5. Cover the plate with one plate sealer and incubate plate at room temperature for 1 hour.
6. Prepare HAMA Tracer antibody working solution by 1:21 fold dilution of the antibody (30196) with the tracer Antibody Diluent (30052). For each strip, it is required to mix 1 mL of the tracer antibody diluent with 50 µL of the tracer antibody in a clean test tube.
7. Remove plate sealer. Aspirate the contents of each well. Wash each well 5 times by dispensing 350 µL of working wash solution into each well and then completely aspirating the contents. Alternatively, an automated microplate washer can be used.
8. Add 100 µL of above diluted HAMA Tracer Antibody working solution to each of the wells.
9. Cover the plate with the plate sealer and incubate plate at room temperature for 30 min.
10. Remove plate sealer. Aspirate the contents of each well. Wash each well 5 times by dispensing 350 µL of working wash solution into each well and then completely aspirating the contents. Alternatively, an automated microplate washer can be used.
11. Add 100 µL of ELISA HRP Substrate into each of the wells.
12. Cover the plate with one plate sealer and also with aluminum foil to avoid exposure to light.
13. Incubate plate at room temperature for 20 min.
14. Remove the aluminum foil and plate sealer. Add 100 µL of ELISA Stop Solution into each of the wells. Mix gently.
15. Read the absorbance at 450 nm within 10 minutes in a microplate reader

NOTE: to reduce the background, one can set the instrument to dual wavelength measurement at 450 nm with background wavelength correction set at 595 nm or 620 nm or 630 nm.
PROCEDURAL NOTES
1. It is recommended that all standards, controls and unknown samples be assayed in duplicate. The average absorbance reading of each duplicate should be used for data reduction and the calculation of results.
2. For patient samples with higher than level 5 standard, it is recommended to measure diluted the specimen with assay buffer at 1:10, 1:100, etc. for a more accurate report.
3. Keep light sensitive reagents in the original amber bottles.
4. Store any unused murine IgG coated strips in the foil Ziploc bag with desiccant to protect from moisture.
5. Careful technique and use of properly calibrated pipetting devices are necessary to ensure reproducibility of the test.
6. Incubation times or temperatures other than those stated in this insert may affect the results.
7. Avoid air bubbles in the microwell as this could result in lower binding efficiency and higher CV% of duplicate reading.
8. All reagents should be mixed gently and thoroughly prior use. Avoid foaming.

INTERPRETATION OF RESULTS
1. Calculate the average absorbance for each pair of duplicate test results.
2. Subtract the average absorbance of the STD 1 (0 ng/mL) from the average absorbance of all other readings to obtain corrected absorbance.
3. The standard curve is generated by the corrected absorbances of all standard levels on the ordinate against the standard concentration on the abscissa using point-to-point or log-log paper. Appropriate computer assisted data reduction programs may also be used for the calculation of results. We recommend using Quadratic curve fit.

The HAMA concentrations for the controls and patient samples are read directly from the standard curve using their respective corrected absorbance. If log-log graphic paper or computer assisted data reduction program utilizing logarithmic transformation are used, sample having corrected absorbance between the 2nd standard and the next highest standard should be calculated by the formula:

\[
\frac{\text{Corrected absorbance (unknown)}}{\text{Corrected Absorbance (2nd STD)}} \times \text{Value of the 2nd STD}
\]
EXAMPLE DATA AND STANDARD CURVE
A typical absorbance data and the resulting standard curve from Human Anti-Mouse Antibody (HAMA) ELISA Assay Kit are represented. This curve should not be used in lieu of standard curve run with each assay.

<table>
<thead>
<tr>
<th>Well I.D.</th>
<th>OD 450 nm Absorbance</th>
<th>Results ng/mL</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Readings</td>
<td>Average</td>
</tr>
<tr>
<td>0 ng/mL</td>
<td>0.051</td>
<td>0.053</td>
</tr>
<tr>
<td></td>
<td>0.054</td>
<td></td>
</tr>
<tr>
<td>40 ng/mL</td>
<td>0.182</td>
<td>0.183</td>
</tr>
<tr>
<td></td>
<td>0.184</td>
<td></td>
</tr>
<tr>
<td>135 ng/mL</td>
<td>0.561</td>
<td>0.556</td>
</tr>
<tr>
<td></td>
<td>0.552</td>
<td></td>
</tr>
<tr>
<td>450 ng/mL</td>
<td>1.737</td>
<td>1.682</td>
</tr>
<tr>
<td></td>
<td>1.627</td>
<td></td>
</tr>
<tr>
<td>1500 ng/mL</td>
<td>3.230</td>
<td>3.183</td>
</tr>
<tr>
<td></td>
<td>3.136</td>
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</tr>
<tr>
<td>Control 1</td>
<td>0.284</td>
<td>0.296</td>
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<tr>
<td></td>
<td>0.309</td>
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<tr>
<td>Control 2</td>
<td>1.166</td>
<td>1.138</td>
</tr>
<tr>
<td></td>
<td>1.109</td>
<td></td>
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</tbody>
</table>

HAMA ELISA Standard Curve
EXPECTED VALUES
One hundred seventy normal adult sera were measured with this HAMA ELISA. One hundred sixty sera showed the OD reading very close to the zero calibrator. The 99% confidence normal cut-off is 25 ng/ml.

It is highly recommend that each laboratory establish its own normal cut off level.

One positive sample with HAMA level of 64 ng/ml was further tested with dilution of this sample in 1:2, 1:4 and 1:8. A linear HAMA dilution result was observed and indicated HAMA specific activity of this sample.

LIMITATION OF THE PROCEDURE
1. Since there is no Gold Standard concentration or international standard available for HAMA measurement, the values of assay standards were established and validated. Results obtained with different assay methods or kits cannot be used interchangeably.
2. For unknown sample value read directly from the assay that is greater than 1500 ng/mL, it is recommend to measure a further diluted sample for more accurate measurement.
3. Bacterial or fungal contamination of serum specimens or reagents, or cross contamination between reagents may cause erroneous results.
4. Water deionized with polyester resins may inactivate the horseradish peroxidase enzyme.

QUALITY CONTROL
To assure the validity of the results each assay should include adequate controls with known HAMA levels. We recommend that all assays include the laboratory’s own HAMA controls in addition to those provided with this kit.

PERFORMANCE CHARACTERISTICS

Sensitivity
The sensitivity of this HAMA ELISA as determined by the 95% confidence limit on 20 duplicate determination of zero standard is about 2 ng/mL.

High Dose “hook” effect
This assay has showed that it did not have any high dose “hook” effect up to 1,000,000 ng/mL.

Precision
The intra-assay precision was validated by measuring one control sample in a single assay with eight-replicate determinations.

<table>
<thead>
<tr>
<th>Mean HAMA Value (ng/mL)</th>
<th>CV (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>51.66</td>
<td>5.1</td>
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</tbody>
</table>

The inter-assay precision is validated by measuring one control sample in duplicate in 6 individual assays.

<table>
<thead>
<tr>
<th>Mean HAMA Value (ng/mL)</th>
<th>CV (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>52.12</td>
<td>5.8</td>
</tr>
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</table>
Linearity
Two HAMA standard levels were diluted with assay buffer and tested. The results of HAMA percent recovery value in pg/mL are as follows:

<table>
<thead>
<tr>
<th>#</th>
<th>DILUTION</th>
<th>OBSERVED VALUE</th>
<th>RECOVERY %</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>Neat</td>
<td>88.51</td>
<td>-</td>
</tr>
<tr>
<td></td>
<td>1:2</td>
<td>44.98</td>
<td>101</td>
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<td></td>
<td>1:4</td>
<td>22.85</td>
<td>103</td>
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<td></td>
<td>1:8</td>
<td>14.14</td>
<td>113</td>
</tr>
<tr>
<td>2</td>
<td>Neat</td>
<td>298.12</td>
<td>-</td>
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<tr>
<td></td>
<td>1:2</td>
<td>141.93</td>
<td>95</td>
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<tr>
<td></td>
<td>1:4</td>
<td>66.78</td>
<td>90</td>
</tr>
<tr>
<td></td>
<td>1:8</td>
<td>37.15</td>
<td>100</td>
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</table>

REFERENCES
**Warranty Information**

Eagle Biosciences, Inc. warrants its Product(s) to operate or perform substantially in conformance with its specifications, as set forth in the accompanying package insert. This warranty is expressly limited to the refund of the price of any defective Product or the replacement of any defective Product with new Product. This warranty applies only when the Buyer gives written notice to the Eagle Biosciences within the expiration period of the Product(s) by the Buyer. In addition, Eagle Biosciences has no obligation to replace Product(s) as result of a) Buyer negligence, fault, or misuse, b) improper use, c) improper storage and handling, d) intentional damage, or e) event of force majeure, acts of God, or accident.

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For further information about this kit, its application or the procedures in this kit, please contact the Technical Service Team at Eagle Biosciences, Inc. at info@eaglebio.com or at 866-411-8023.