MB440_IFU_EN_V2601

NZYSupreme qPCR Green Master Mix (2x), ROX plus

Catalogue number	Presentation
MB44001	2 x 1 mL (200 rxns of 20 µL)
MB44002	5 x 1 mL (500 rxns of 20 µL)
MB44003	20 x 1 mL (2000 rxns of 20 µL)
MB44005	1 x 50 mL (5000 rxns of 20 µL)

Description

NZYSupreme qPCR Green Master Mix (2x), ROX plus is an optimized and highly efficient reaction mixture developed for real-time PCR. This master mix was engineered with a hot-start enzyme control mechanism to provide the highest detection sensitivity. In addition, the latest developments in PCR enhancers have been incorporated in the NZYSupreme qPCR Green Master Mix, ROX plus, including buffer chemistry and highly robust engineered enzymes. These combinations guarantee that NZYSupreme qPCR Green Master Mix (2x), ROX plus delivers ultra-sensitivity coupled with highly reproducible and fast real-time PCR protocols. It was designed to amplify targets for accurate gene expression analysis. The master mix is provided as a 2x reaction mixture that contains all components necessary for real-time PCR, including a green intercalating dye, dNTPs, stabilizers and enhancers. The presence of ROX plus reference dye in the master mix enables to increase confidence in data analysis, since it allows to normalize non-PCR-related fluctuations in fluorescence. Despite most real-time PCR instruments that can read ROX plus dye allow users to run experiments and analyze data without ROX plus, the inclusion of this internal passive reference dye prevents data misinterpretation and allows to detect and diagnose errors. NZYSupreme qPCR Green Master Mix (2x), ROX plus is ready-to-use and only requires primers and DNA template addition. It is optimized for intercalating green dye detection on different instruments.

Shipping & Storage Conditions

The product can be shipped in a range of temperatures from dry ice to blue ice. Upon arrival, all components should be stored at -85 °C to -15 °C in a constant temperature freezer to guarantee maximal shelf life. Minimize the number of freeze-thaw cycles by storing it in working aliquots. The master mix remains stable at 2-8 °C for up to 5 months. Do not expose the master mix to direct sunlight. The product will remain stable till the expiry date if stored as specified.

Components

COMPONENT	SKU	TUBES/BOTTLES	VOLUME
NZYSupreme qPCR Green Master Mix (2x), ROX plus	**MB44001**	1	1 mL
	MB44002	5	1 mL
	MB44003	20	1 mL
	MB44005	1	50 mL

Note: Consider preparing multiple aliquots of the master mix to reduce freeze/thaw cycles and minimize the risk of contamination.

Specifications

Compatibility with real-time PCR instruments

NZYSupreme qPCR Green Master Mix (2x), ROX plus is compatible with instruments that measure the passive reference signal. However, it is also compatible with instruments that do not require a passive reference signal for data normalization. It was optimized to be compatible with the following real-time PCR instruments:

Applied Biosystems: 7000; 7300; 7700; 7900; 7900HT; 7900HT FAST; StepOne™ & StepOne™plus

Standard Protocol

Recommendations before starting

- Nucleic acid manipulation: Stringent precautionary measures must be imposed to mitigate the risk of carry-over contamination of DNA. We recommend using DNase-free plasticware/reagents and working in a DNase-free area (Nucleases & Nucleic Acid Cleaner, Cat. No. MB48301, or DNA & RNA Cleaner, Cat. No. MB46201, can help remove DNases from surfaces and materials).
- Handling instructions: To help prevent any carry-over DNA contamination, you should assign independent areas for reaction set-up, PCR amplification and any post-PCR gel analysis. Any tubes containing amplified PCR product mustn't be opened in the PCR set-up area. Use sterile filtered tips. Minimize exposure by keeping reaction and components capped whenever possible.

FOR REFERENCE USE ONLY

- Controls: For verification of the absence of contamination, prepare a mixture sample without a DNA template (negative control). Additionally, include a positive control to serve as a reference for ensuring the correct functioning of the qPCR reaction and detection system. The positive control should exhibit the expected amplification and/or fluorescence signal, confirming the assay's ability to accurately detect the target sequence. To avoid incorrect results caused by unwanted fluorescent substances, consider adding a No-Amplification Control tube. This tube should have the sample but not the enzyme master mix. Elevated fluorescence in the No-Amplification Control compared to the No-Template Control suggests potential fluorescent contaminants in either the sample or the thermal cycler's heat block.
- Replicates: It is highly recommended to perform replicates of each reaction; we recommend performing four replicates or at least three.

Procedure

The following protocol serves as a general guideline and a starting point for any qPCR procedure. Optimal reaction conditions (*e.g*. incubation times, temperatures and template concentration) may vary and, in particular conditions, may require further optimization.

1. Thaw the master mix at room temperature or on ice. Mix the master mix thoroughly by flicking the tube and inverting.
   Note: *To ensure optimal performance, please make sure all mix components are thawed and resuspended/homogenized before use. Before pipetting, mix vigorously the master mix by inverting the tube and then spin down.*
2. In a clean reaction setup area, prepare the qPCR reaction mixture according to the table below (please notice that the given volumes are based on a standard 20 µL final reaction mix and can be scale adjusted):
   Note 1: *If setting up more than one reaction, prepare a reaction mixture volume 10% greater than the total required for the number of reactions to be performed. Include sufficient reactions for the negative and positive controls.*
   Note 2: *If necessary, prepare a No-Amplification Control by adding nuclease-free water instead of the qPCR master mix.*
   Note 3: *We recommend performing replicates of all reactions.*

	1 REACTION VOLUME	FINAL CONCENTRATION
NZYSupreme qPCR Green Master Mix (2x), ROX plus (1)	10 µL	1×
10 µM forward primer	0.8 µL	400 nM (2)
10 µM reverse primer	0.8 µL	400 nM (2)
FINAL VOLUME =	up to 11.6 µL (3)	-

(1) Make sure all mix components are thawed and resuspended/homogenized before use, please refer to the note above (step 1).
(2) Refer to the section of "Technical Notes" below for more details about primers final concentrations in the reaction.
(3) If using smaller volumes for the primers, supplement the volume up to 11.6 µL with Nuclease-free Water.

3. Gently mix and centrifuge briefly to spin down the contents.
4. Pipette 11.6 µL of the Mix into each well, according to your experimental plate/strip/tube configuration.
5. Pipette template (samples and controls):
   a. Add up to 8.4 µL of DNA sample/positive control into each respective well. If using less volume than 8.4 µL, supplement with Nuclease-free Water up to 8.4 µL. In total, the final volume of the reaction will be 20 µL.
   b. Add up to 8.4 µL of Nuclease-free Water for the negative control to achive a final volume of 20 µL.

Note: *To avoid cross-contamination, we strongly recommend pipetting the template at last, preferably in a work separate area.*

6. Cover and seal the plate/strip/tube with appropriate caps or optical adhesive film before proceeding with the real-time PCR detection steps.
7. Centrifuge briefly to spin down the contents and eliminate any air bubbles from the reaction mixtures.
8. Place the reaction plate/strip/tube within the real-time PCR instrument and run the general protocol defined below. These conditions might be adapted to suit your specific needs, within sensible limits.

Suggested thermal cycling conditions

NZYSupreme qPCR Green Master Mix (2x), ROX plus was optimized for the amplification of DNA fragments up to 200 bp under different real-time PCR cycling conditions. The table below displays a standard qPCR setup optimized on several platforms. However, these conditions may be adapted to suit different equipment-specific protocols.

CYCLES	TEMP.	TIME	CYCLE STEP
1	95 °C	2 - 5 min (*)	Polymerase activation
40	95 °C 60 °C	5 sec 15 - 30 sec	Denaturation Annealing/Extension

(*) 2 min for cDNA, up to 3 or 5 min for genomic DNA.

Melting curve analysis: At the end of the qPCR run, it is highly recommended to perform a melting curve. A melt curve performed after qPCR cycling with an intercalating dye will typically produce a single distinct peak. This indicates that the amplified double-stranded DNA products are a single discrete species. The presence of multiple DNA species in the same reaction produces multiple peaks in the melt curve, typically indicating the presence of contaminating or off-target amplification products. Follow the manufacturer's instructions for the real-time PCR instrument for melting curve analysis

FOR REFERENCE USE ONLY

Testing and Ct values

When comparing NZYSupreme qPCR Green Master Mix (2x), ROX plus with a mix from another supplier we strongly recommend amplifying from a 10-fold template dilution series. Loss of detection at low template concentration is the only direct measurement of sensitivity. An early Ct value is not an indication of good sensitivity, but rather an indication of speed.

Technical Notes

Primers: The specific amplification, yield and overall efficiency of any real-time PCR can be critically affected by the sequence and concentration of the primers, as well as by the amplicon length. We strongly recommend considering the following points when designing and running your real-time PCR experiment:

- Primers should have a melting temperature (Tm) of approximately 58-62 °C;
- The fragment to amplify should be between 70-200 bp in length and not superior to 400 bp;
- Final primer concentrations of 400 nM are suitable for *green* reactions. However, to determine the optimal concentration we recommend titrating in the range 100-1000 nM. Forward and reverse primers concentration should be equimolar;
- Design intron spanning primers when amplifying from cDNA (to avoid gDNA amplification).

Template: The DNA template must be purified and devoid of contamination by PCR inhibitors (*e.g.* EDTA). The DNA template must be purified and concentrated according to conventional nucleic acid clean-up procedures (NZYGelpure, MB011). The recommended amount of template is dependent upon the source of DNA used. Please consider the following points when selecting genomic DNA or cDNA templates:

- **Genomic DNA:** use up to 1 µg of genomic DNA in a single PCR. We recommend using the NZY Tissue gDNA Isolation kit (Cat. No. MB135) for high yield and purity from both prokaryotic and eukaryotic sources.
- **cDNA:** the optimal amount of cDNA to use in a single PCR depends upon the copy number of the target gene; we suggest using 100 ng cDNA per reaction. However, this amount may be adjusted to a more appropriate concentration. We suggest using the NZY First-Strand cDNA Synthesis Kit (Cat. No. MB125) for reverse transcription of purified RNA. To obtain a high yield of highly purified RNA we suggest using NZY Total RNA Isolation Kit (Cat. No. MB134).

$MgCl_2$: It is not necessary to supplement the reaction mixture with $MgCl_2$ as the NZYSupreme qPCR Green Master Mix (2x), ROX plus already contains an optimized concentration of $MgCl_2$.

PCR controls: The reliability of the data may be affected by the presence of contaminating DNA. It is strongly recommended to include a no-template control reaction in the qPCR design, replacing template DNA/cDNA with nuclease-free PCR-grade water (DEPC-treated Water, Cat. No. MB43701).

Green intercalating dye: NZYSupreme qPCR Green Master Mix (2x), ROX plus contains a non-specific double-strand DNA-binding dye, that will bind to all dsDNA fragments present in the reaction. Upon binding to DNA, it emits green fluorescence (λ = 520 nm) while showing no detectable inhibition to the PCR reaction.

Quality control assays

Genomic DNA contamination

The product must comply with internal standards of DNA contamination as evaluated through real-time PCR.

Nuclease assays

To test for DNase contamination, 0.2-0.3 µg of pNZY28 plasmid DNA are incubated with the master mix for 14-16 h at 37 °C. To test for RNase contamination, 1 µg of RNA is incubated with the master mix for 1 h at 37 °C. Following incubation, the nucleic acids are visualized on a GreenSafe-stained agarose gel. There must be no visible nicking or cutting of the nucleic acids.

Functional assay

NZYSupreme qPCR Green Master Mix (2x), ROX plus is extensively tested for activity, processivity, efficiency, sensitivity and heat activation.

This master mix is manufactured under stringent quality standards and complies with ISO 9001 and ISO 13485 certifications for research and diagnostic-grade reagents.

FOR REFERENCE USE ONLY