Triiodothyronine (T3) ELISA Assay Kit

Catalog Number: T3T31-K01 (1 x 96 wells)
For Research Use Only. Not for use in diagnostic procedures.

v. 10 (20 FEB 24)

EAGLE BIOSCIENCES, INC.
20A NW Blvd., Suite 112, Nashua, NH 03063
Phone: 617-419-2019Fax: 617-419-1110
WWW.EAGLEBIO.COM

INTENDED USE

The Eagle Biosciences Triiodothyronine (T3) ELISA Assay Kit (enzyme-linked immunoassay kit) is intended for the quantitative measurement of triiodothyronine (T3) in human serum. The Eagle Biosciences Triiodothyronine (f3) ELISA Assay Kit is for research use only and not to be used in diagnostic procedures.

INTRODUCTION

Triiodothyronine (T3) and thyroxine (T4) are the two active thyroid hormones found in the blood stream. About 80% of T3 is produced by the deiodination of T4 in the peripheral tissue and the other 20% is produced directly from the thyroid gland. T3 is transported through the peripheral blood stream bound to serum proteins, namely thyroxine binding globulin, thyroid binding prealbumin and albumin. About 0.3% of the total T3 is unbound and is therefore considered the free fraction.

T3 has an influence on oxygen consumption and heat production in virtually all tissues. The hormone also plays a critical role in growth, development and sexual maturation of growing organisms.

T3 is one parameter used in the clinical diagnosis and differentiation of thyroid disease, particularly hyperthyroidism. In most hyperthyroid patients, both serum T3 and serum T4 levels are elevated. Serum T3 levels are a sensitive indicator of the impending hyperthyroid state often preceding elevated T4 and free thyroxine index values. Serum T3 levels are clinically significant in both the diagnosis of thyroid disease and in the detection of T3-thyrotoxicosis. However, it has been demonstrated that T3 levels may be affected by a number of medications, acute and chronic stress, and a variety of acute and chronic nonthyroidal illnesses. It is therefore necessary to differentiate those results that are due to thyroid dysfunction from those related to non-thyroidal diseases.

PRINCIPLE OF THE ASSAY

The T3 ELISA is a competitive immunoassay. Competition occurs between T3 present in calibrators, controls, specimen samples and an enzyme-labelled antigen (HRP conjugate) for a limited number of anti-T3 antibody binding sites on the microplate wells. After a washing step that removes unbound materials, TMB substrate (enzyme substrate) is added which reacts with HRP to form a blue-colored product that is inversely proportional to the amount of T3 present. Following an incubation, the enzymatic reaction is terminated by the addition of the stopping solution, converting the color from blue to yellow. The absorbance is measured on a microtiter plate reader at 450 nm. A set of calibrators is used to plot a calibrator curve from which the amount of T3 in specimens samples and controls can be directly read.

PROCEDURAL CAUTIONS AND WARNINGS

1. This kit is for use by trained laboratory personnel (professional use only). For laboratory in vitro use only.
2. Practice good laboratory practices when handling kit reagents and specimens. This includes:
    a. Do not pipette by mouth.
    b. Do not smoke, Drink, or eat in areas where specimens or kit reagents are handled.
    c. Wear protective clothing and disposable gloves.
    d. Wash hands thoroughly after performing the test.
    e. Avoid contact with the eyes, use safety glasses; in case of contact with eyes, flush eyes with water immediately and contact a doctor.

3. Users should have a thorough understanding of this protocol for the successful use of this kit. Reliable performance will only be attained by strict and careful adherence to the instructions provided.
4. Do not use this kit beyond the expiry date stated on the label.
5. If the kit reagents are visibly damaged, do not use the test kit.
6. Do not use kit components from different kit lots within a test and do not use any component beyond the expiration date printed on the label.
7. All kit reagents and specimens must be brought to room temperature and mixed gently but thoroughly before use. Avoid repeated freezing and thawing of specimens.
8. When the use of water is specified for dilution or reconstitution, use deionized or distilled water.
9. Immediately after use, each individual component of the kit must be returned to the recommended storage temperature stated on the label.
10. A calibrator curve must be established for every run.
11. It is recommended to all customers to prepare their own control materials or serum pools which should be included in every run at a high and low level for assessing the reliability of results.
12. The controls (included in kit) must be included in every run and their results must fall within the ranges stated in the quality control certificate; a failed control result might indicate improper reagent storage.
13. When dispensing the substrate and stopping solutions, do not use pipettes in which these liquids will come into contact with any metal parts.
14. The TMB Substrate is sensitive to light and should remain colorless if properly stored. Instability or contamination may be indicated by the development of a blue color, in which case it should not be used.
15. Do not use grossly hemolyzed, grossly lipemic, icteric or improperly stored serum.
16. Samples or controls containing azide or thimerosal are not compatible with this kit, they may lead to false results.
17. Samples values above the measuring range of the kit may be reported as >10 ng/mL. If further dilution and retesting is required, only Calibrator A may be used to dilute serum samples. The use of any other reagent may lead to false results.
18. Avoid microbial contamination of reagents.
19. To prevent the contamination of reagents, use a new disposable pipette tip for dispensing each reagent, sample, calibrator, and control.
20. To prevent contamination of reagents, do not pour reagents back into the original containers.
21. Kit reagents must be regarded as hazardous waste and disposed of according to local and/or national regulations.
22. Consumables used with the kit that are potentially biohazardous (e.g., pipette tips, bottles or containers containing human materials) must be handled according to biosafety practices to minimize the risk of infection and disposed of according to local and/or national regulations relating to biohazardous waste.
23. This kit contains 1 M sulfuric acid in the stopping solution component. Do not combine acid with waste material containing sodium azide or sodium hypochlorite.
24. The use of safety glasses, and disposable plastic, is strongly recommended when manipulating biohazardous or bio-contaminated solutions.
25. Proper calibration of the equipment used with the test, such as the pipettes and absorbance microplate reader, is required.
26. If a microplate shaker is required for the assay procedure, the type and speed of shaker required is stated in the REAGENTS AND EQUIPMENT NEEDED BUT NOT PROVIDED section. Both the type and speed of saker used can influence the optical

densities and test results. If a different type of shake and/or speed is used, the user is responsible for validating the performance of the kit.
27. Do not reuse the microplate wells, they are for SINGLE USE only.
28. To avoid condensation within the microplate wells in humid environments, do not open the pouch containing the microplate until it has reached room temperature.
29. Any serious incident that has occurred in relation to the device shall be reported to the manufacturer and the competent authority of the Member State in which the user and/or the participant is established.

SAFETY CAUTIONS AND WARNINGS

Biohazards
The reagents should be considered a potential biohazard and handled with the same precautions applied to blood specimens. All human specimens should be considered a potential biohazard and handled as if capable of transmitting infections and in accordance with good laboratory practices.

Chemical Hazards
Avoid direct contact with any of the kit reagents. Specifically, avoid contact with the TMB Substrate (contains tetramethylbenzidine) and Stopping Solution (contains Sulfuric Acid). If contacted with any of these reagents, wash with plenty of water and refer to SDS for additional information.

SPECIMEN COLLECTION, STORAGE, AND PRE-TREATMENT

Specimen Collection & Storage
Approximately 0.15 mL of serum is required per duplicate determination. Collect 4–5 mL of venous blood into an appropriately labelled tube and allow it to clot. Centrifuge at room temperature and carefully transfer the serum into a new storage tube or container. Serum samples may be stored at 2-8°C for up to 24 hours or at -10°C or lower if the analyses are to be done at a later date.
Consider all human specimens as possible biohazardous materials and take appropriate precautions when handling.

Specimen Pre-Treatment
Specimen pre-treatment is not required.

REAGENTS AND EQUIPMENT NEEDED BUT NOT PROVIDED

1. Calibrated single-channel pipette to dispense 50 µL
2. Calibrated multi-channel pipette to dispense 50 µL, 100 µL and 150 µL
3. Calibrated multi-channel pipettes to dispense 350 µL (if washing manually)
4. Automatic microplate washer (recommended)
5. Disposable pipette tips
6. Distilled or deionized water
7. Calibrated absorbance microplate reader with a 450 nm filter and an upper OD limit of 3.0 or greater.

REAGENTS PROVIDED

1. **Microplate**

Contents:	One anti-T3 polyclonal antibody-coated 96-well (12x8) microplate in a resealable pouch with desiccant.
Format:	Ready to use
Storage:	2-8°C
Stability:	Unopened: Stable until the expiry date printed on the label. After Opening: Stable for four weeks.

2. **HRP Conjugate Concentration**

Contents:	One bottle containing T3-Horse Radish Peroxidase (HRP) conjugate in a protein-based buffer with a non-mercury preservative
Format:	Ready to use
Volume:	15mL/bottle
Storage:	2-8°C
Stability:	Unopened: Stable until the expiry date printed on the label. After Opening: Stable for four weeks

3. **Calibrator A – E**

Contents:	Five bottles of calibrator containing specified T3 concentrations. Protein-based buffer with a non-mercury preservative. Prepared by spiking buffer with define quantities of T3. Listed below are approximate concentrations, please refer to vial labels for exact concentrations. *Concentrations: 0, 0.3, 1, 3, 10 ng/mL*
Format:	Ready to use
Volume:	Calibrator A: 2.0 mL/bottle
	Calibrator B-E: 1.0 mL/bottle
Storage:	2 – 8°C
Stability:	Unopened: Stable until the expiry date printed on label. After opening: Stable for four weeks.

4. **Control 1 – 2**

Contents:	Two bottles of control containing different T3 concentrations. Protein-based buffer with a non-mercury preservative. Prepared by spiking buffer with defined quantities of T3. Refer to the QC certificate for the target values and acceptable ranges.
Format:	Ready to Use
Volume:	1.0 mL/bottle
Storage:	2 - 8°C
Stability:	Unopened: Stable until the expiry date printed on the label. After Opening: Stable for four weeks.

5. **TMB Substrate**

Contents:	One bottle containing tetramethylbenzidine and hydrogen peroxide in a non-DMF or DMSO containing buffer.
Format:	Ready to Use
Volume:	18 mL/bottle
Storage:	2 - 8°C
Stability:	Unopened: Stale until the expiry printed on the label.
	After Opened: Stable for four weeks.

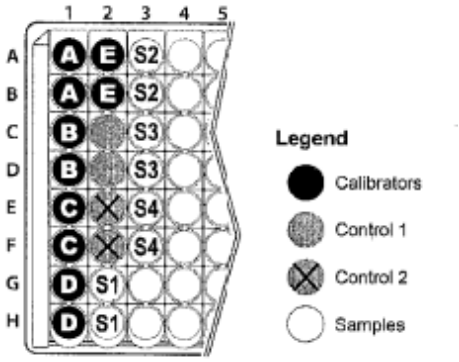
6. **Stopping Solution**

Contents:	One bottle containing 1M sulfuric acid
Format:	Ready to Use
Volume:	6mL/bottle
Storage:	2 - 8°C
Safety:	Refer to product SDS.

7. **Wash Buffer Concentrate**

Contents:	One bottle containing buffer with a non-ionic detergent and a non-mercury preservative.
Format:	Concentrated; Requires Preparation
Volume:	50 mL/bottle
Storage:	2 - 8°C
Stability:	Unopened: Stable until the expiry date printed on the label. After Opening: Stable for four weeks. Following Preparation: The wash buffer working solution is stable for 2 weeks following preparation, assuming Good Laboratory Practices are adhered to. To prevent microbial growth, prepare the wash buffer working solution in a clean container and store under refrigerated conditions (2-8°C) when not in use.
Preparation:	Dilute 1:10 in distilled or deionized water before use. If the whole microplate is to be used dilute 50 mL of the wash buffer concentrate in 450 mL of distilled or deionized water.

RECOMMENDED ASSAY LAYOUT



ASSAY PROCEDURE

Specimen Pretreatment: None.

All kit components, controls, and specimen samples must reach room temperature before use. Calibrators, controls and specimen samples should be assayed in duplicate. Once the procedure has been started, all steps should be completed without interruption.

1. After all kit components have reached room temperature, **mix** gently by inversion.
2. Prepare the HRP Conjugate Working Solution and Wash Buffer Working Solution
3. (See section Reagents Provided, HRP Conjugate Concentration, Wash Buffer Concentrate).
4. Plan the microplate wells to be used for calibrators, controls, and samples. See Recommended Assay Layout. Remove the strips from the microplate frame that will not be used and place them in the bag with desiccant. Reseal the bag with the unused strips and return it to the refrigerator.
5. Pipette 50 µL of each calibrator, control, and specimen sample into assigned wells.
6. Pipette 100 µL of the HRP Conjugate into each well (the use of a multi-channel pipette is recommended).
7. Wash the microplate wells with an automatic microplate washer (preferred) or manually as stated below.
    a. Automatic: Using an automatic microplate washer, perform a 3-cycle wash using 350 µL /well of Wash Buffer Working Solution (3 x 300 µL). One cycle consists of aspirating all wells then filling each well with 300 µL of Wash Buffer Working Solution. After the final wash cycle, aspirate all wells and then tap the microplate firmly against absorbent paper to remove any residual liquid.
    b. Manually: For manual washing, perform a 3-cycle wash using 350 µL /well of Wash Buffer Working Solution (3 x 300 µL). One cycle consists of aspirating all wells by briskly emptying the contents of the wells over a waster container, then pipetting 300 µL of Wash Buffer Working Solution into each well using a multi-channel pipette. After the final wash cycle, aspirate all wells by briskly emptying the contents over a waste container and then tap the microplate firmly against absorbent paper to remove any residual liquid.
8. Pipette 150 µL of TMB Substrate into each well (the use of a multi-channel pipette is recommended).
9. Incubate the microplate for 15 minutes at room temperature.
10. Pipette 50 µL of Stopping Solution into each well (the use of a multi-channel pipette is recommended) in the same order and speed as was used for the addition of TMB Substrate. Gently tap the microplate frame to mix the contents of the wells.
11. Measure the optical density (absorbance) in the microplate wells using an absorbance microplate reader set to 450 nm, within 20 minutes after addition of the Stopping Solution.

CALCULATIONS

1. Calculate the mean optical density of each calibrator, control, and sample duplicate.
2. Use a 4-parameter or 5-parameter curve fit with immunoassay software to generate a calibrator curve.
3. The immunoassay software will calculate the concentrations of the controls and specimen samples using the mean optical density values and the calibrator curve.

4. If a sample reads more than 10 ng/mL and needs to be diluted and retested, then dilute with calibrator A not more than 1:5. The result obtained must be multiplied by the dilution factor.

QUALITY CONTROL

When assessing the validity of the test results, the following criteria should be evaluated:

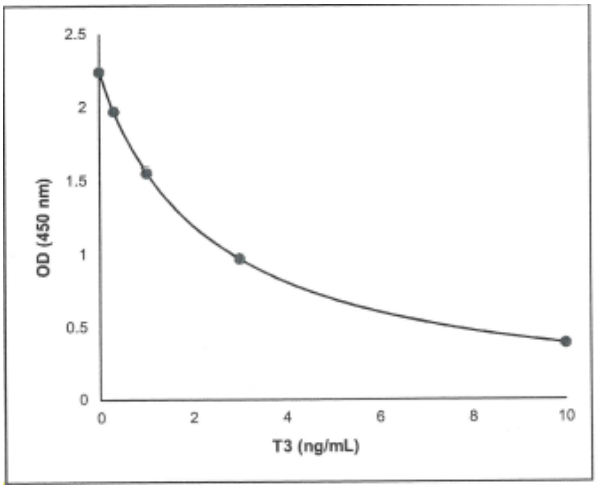
1. The calibrator A mean optical density meets the acceptable range as stated in the QC Certificate.
2. The calibrator with the highest concentration meets the % binding acceptable ranges as stated in the QC Certificate. % Binding = (OD of calibrator / OD of calibrator A) x 100.
3. The values obtained for the kit controls are withing the acceptable ranges as stated in the QC certificate.
4. The results of any external controls that were used meet the acceptable ranges.

TYPICAL TABULATED DATA

Sample data only. Do not use to calculate results.

Calibrator	Mean OD (450 nm)	% Binding	Value (ng/mL)
A	2.239	100	0
B	1.969	88	0.3
C	1.549	69	1
D	0.966	43	3
E	0.383	17	10
Unknown	1.512	-	1.1

TYPICAL CALIBRATOR CURVE



PERFORMANCE CHARACTERISTICS

Sensitivity

The analytical sensitivity study was performed according to the CLSI EP17-A2 guideline. The Limit of Background (LoB), Limit of Detection (LoD), and the Limit of Quantitation (LoQ) are summarized in the table below.

Parameter	T3 (ng/mL)
LoB	0.082
LoD	0.153
LoQ	0.280

Specificity

The following compounds were tested for cross-reactivity with the Direct fT3 ELISA kit with T3 cross-reacting at 100%.

Compound	% Cross Reactivity
3,3',5-Triiodo-L-thryonine (T3)	100
3,3',5'-Triiodo-L-thryonine (Reverse T3)	2.03
3,5-Diiodo-L-thryonine (3,5-T2)	1.98
3,5-Diiodo-L-tyrosine dihydrate	<0.001
Phenytoin (Phenytoin-D10)	0.003
D-Thyroxine	0.64
L-Thyroxine	0.60

INTERFERENCES

An interference study was performed according to the CLSI EP07 guideline. No significant interference was observed for concentrations of up to 10 g/L hemoglobin, 15 mg/dL bilirubin conjugated, 30 mg/dL bilirubin unconjugated, 15 mg/mL triglycerides, 1.8 µg/mL HAMAS, 3.6 µg/mL Biotin and 1688 IU/mL Rheumatoid Factor.

PRECISION

The precision study was performed according to the CLSI EP5-A3 guideline.
The experimental protocol used a nested components-of-variance design (a 3 x 1 x 5 x 2 x 4 design) with 5 testing days, an automated machine and 2 operators, 1 kit lot, six runs per testing day (two runs per operator or automated machine), and four replicate measurements per run for each sample (total of 120 results per sample). The results were analyzed with a two-way nested ANOVA and summarized in the table below.

Sample	Mean (ng/mL)	SD/CV %	Repeatability	Reproducibility	Within Condition	Between Condition	Between Replicate	Between Day
		SD	0.08	0.11	0.11	0.03	0.05	0.06
1	0.809	CV	9.9	13.9	13.4	3.7	5.8	7.1
2	2.147	SD	0.13	0.21	0.21	0.05	0.04	0.16
		CV	5.9	9.9	9.6	2.4	1.6	7.4
3	4.075	SD	0.18	0.39	0.32	0.23	0.15	0.22
		CV	4.4	9.7	7.9	5.6	3.6	5.5
4	1.110	SD	0.08	0.12	0.12	0.04	0.06	0.06
		CV	7.3	11.2	10.6	3.6	5.3	5.5
		SD	0.13	0.17	0.14	0.09	0.00	0.05
5	2.100	CV	6.0	7.9	6.5	4.5	0.0	2.6
6	3.335	SD	0.14	0.22	0.17	0.14	0.07	0.07
		CV	4.1	6.6	5.0	4.3	2.1	2.1
7	0.410	SD	0.6	0.07	0.07	0.02	0.00	0.04
		CV	14.0	17.6	17.2	4.12	0.0	9.9
8	5.962	SD	0.20	0.40	0.29	0.28	0.10	0.18
		CV	3.3	6.7	4.8	4.6	1.7	3.1
9	1.077	SD	0.10	0.14	0.13	0.04	0.05	0.07
		CV	9.1	12.9	12.2	4.2	4.6	6.7
10	1.110	SD	0.09	0.11	0.10	0.05	0.00	0.05
		CV	8.0	10.1	9.0	4.5	0.2	4.1

Linearity

The linearity study was according to the CLSI EP06-A guideline using three human serum samples covering the range of the assay. The samples were diluted in calibrator A at several equidistant concentration levels and the regression equation of the results (y) compared to the concentration (x) predicted from the dilution factor was:

y = 0.99x + 0.17

r = 0.99

The statistical analysis shows that the assay is sufficiently linear up to a 1:5 dilution when using calibrator A as the diluent

RECOVERY

Sample	Observed Result	Expected Result	Recovery %
1 Unspiked	1.028	-	-
+1 ng/mL	1.027	1.014	101.3
+3 ng/mL	2.052	2.014	101.9
+10 ng/mL	5.266	5.514	95.5
2 Unspiked	1.018	-	96.3
+1 ng/mL	0.972	1.009	92.3
+3 ng/mL	1.854	2.009	88.9
+10 ng/mL	4.899	5.509	
3 Unspiked	1.285	-	-
+1 ng/mL	1.019	1.143	89.2
+3 ng/mL	2.115	2.143	98.7
+10 ng/mL	4.903	5.643	86.9
4 Unspiked	1.109	-	-
+1 ng/mL	0.887	1.055	84.1

+3 ng/mL	1.772	2.055	86.2
+10 ng/mL	4.838	5.555	87.1

Reference Ranges

As for all clinical assays each laboratory should collect data and establish their own range of expected normal values.

Group	Range (ng/mL)
Healthy Normal Males and Females	0.76 – 1.66

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10. https://www.uofmhealth.org/health-library/hw27377
11. https://labtestsonline.org/testslphenytoin

WARRANTY INFORMATION

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For further information about this kit, its application or the procedures in this kit, please contact the Technical Service Team at Eagle Biosciences, Inc. at info@eaglebio.com or at 866-411-8023.