



DCM018-13  
Ed. 01/2019

# URINARY CORTISOL ELISA

Direct immunoenzymatic determination of free Cortisol in urine

for routine analysis  
for research use only

IVD



See external label

LOT

2°C 8°C



Σ = 96 tests

REF DKO018

## INTENDED USE

The Eagle Biosciences Urinary Cortisol ELISA Assay Kit is a competitive immunoenzymatic colorimetric method for quantitative determination of free Cortisol concentration in Urine. Urinary Cortisol ELISA kit is for research use only and is not intended for diagnostic procedures.

### 1. CLINICAL SIGNIFICANCE

Cortisol is a steroid hormone released from the adrenal cortex in response to an hormone called ACTH (produced by the pituitary gland), it is involved in the response to stress; it increases blood pressure, blood sugar levels, may cause infertility in women, and suppresses the immune system.

Cortisol acts through specific intracellular receptors and has effects in numerous physiologic systems, including immune function, glucose-counter regulation, vascular tone, substrate utilization and bone metabolism. Cortisol is excreted primarily in urine in an unbound (free) form.

Cortisol is bound, in plasma, from corticosteroid-binding globulin (CBG, transcortin), with high affinity, and from albumin. Only free cortisol is available to most receptors.

These normal endogenous functions are the basis for the physiological consequences of chronic stress - prolonged cortisol secretion causes muscle wastage, hyperglycaemia, and suppresses immune / inflammatory responses. The same consequences arise from long-term use of glucocorticoid drugs.

The free cortisol fraction represents the metabolically active cortisol. In normal conditions, less than 1% it comes excrete in urines. In pathological conditions (syndrome of Cushing) the levels of free urinary cortisol are elevated, because the CBG don't bound the plasmatic cortisol in excess and it was removed with urines.

During pregnancy or estrogen treatment an increase of plasmatic cortisol caused by an increment of the production of the transport protein, but the levels of free urinary cortisol results normal to indicate a correct surrenic functionality.

This test is very useful to estimate the real surrenic function, because is dose the free cortisol, it is the metabolically active form. Moreover the measurement of free urinary cortisol is the better parameter for the diagnosis of the Cushing's syndrome.

## 2. PRINCIPLE

The Cortisol (antigen) in the sample competes with the antigenic Cortisol conjugated with horseradish peroxidase (HRP) for binding to the limited number of antibodies anti Cortisol coated on the microplate (solid phase).

After incubation, the bound/free separation is performed by a simple solid-phase washing.

Then, the enzyme HRP in the bound-fraction reacts with the Substrate (H<sub>2</sub>O<sub>2</sub>) and the TMB Substrate and develops a blue color that changes into yellow when the Stop Solution (H<sub>2</sub>SO<sub>4</sub>) is added.

The colour intensity is inversely proportional to the Cortisol concentration of in the sample.

Cortisol concentration in the sample is calculated through a calibration curve.

## 3. REAGENTS, MATERIALS AND INSTRUMENTATION

### 3.1. Reagents and materials supplied in the kit

#### 1. Calibrators (5 vials)

CAL0	(4 mL)	REF DCE002/1806-0
CAL1	(1 mL)	REF DCE002/1807-0
CAL2	(1 mL)	REF DCE002/1808-0
CAL3	(1 mL)	REF DCE002/1809-0
CAL4	(1 mL)	REF DCE002/1810-0

#### 2. Controls (2 vial, 1 mL each, ready to use)

Low Control	REF DCE045/1801-0
High Control	REF DCE045/1802-0

#### 3. Conjugate (1 vial, 33 mL)

Cortisol conjugated with horseradish peroxidase (HRP)  
REF DCE002/1802-0

#### 4. Coated Microplate (1 breakable microplate)

Anti Cortisol antibody adsorbed on the microplate  
REF DCE002/1803-0

#### 5. TMB Substrate (1 vial, 15 mL)

H<sub>2</sub>O<sub>2</sub>-TMB (0.26 g/L) (avoid any skin contact)  
REF DCE004-0

#### 6. Stop Solution (1 vial, 15 mL)

Sulphuric acid 0.15 mol/L (avoid any skin contact)  
REF DCE005-0

#### 7. 10X Conc. Wash Solution (1 vial, 50 mL)

Phosphate buffer 0.2M, Proclin < 0,0015%  
REF DCE054-0

### 3.2. Reagents necessary not supplied

Distilled water

### 3.3. Auxiliary materials and instrumentation

Automatic dispenser

Microplates reader (450 nm, 620-630 nm)

#### Note

Store all reagents at 2-8°C in the dark.

Open the bag of reagent 4 (Coated Microplate) only when it is at room temperature and close it immediately after use; once opened, the microplate is stable until expiry date of the kit.

## 4. WARNINGS

- This Urinary Cortisol ELISA Assay Kit is intended for research use only by professional persons only. Not for internal or external use in Humans or Animals.
- Use appropriate personal protective equipment while working with the reagents provided.
- Follow Good Laboratory Practice (GLP) for handling blood products.
- Material of animal origin used in the preparation of the kit has been obtained from animals certified as healthy and the bovine protein has been obtained from countries not infected by BSE, but these materials should be handled as potentially infectious.
- Some reagents of the Urinary Cortisol ELISA Assay Kit contain small amounts of Proclin 300<sup>R</sup> as preservative. Avoid the contact with skin or mucosa.
- The TMB Substrate contains an irritant, which may be harmful if inhaled, ingested or absorbed through the skin. To prevent injury, avoid inhalation, ingestion or contact with skin and eyes.
- The Stop Solution consists of a diluted sulphuric acid solution. Sulphuric acid is poisonous and corrosive and can be toxic if ingested. To prevent chemical burns, avoid contact with skin and eyes.
- Avoid the exposure of reagent TMB/H<sub>2</sub>O<sub>2</sub> to directed sunlight, metals or oxidants. Do not freeze the solution.
- This method allows the determination of Cortisol from 0,47 ng/mL (LOD) to 200 ng/mL.
- The clinical significance of the Cortisol determination can be invalidated if the patient was treated with corticosteroids or natural or syntetic steroids.

## 5. PRECAUTIONS

- Please adhere strictly to the sequence of pipetting steps provided in this protocol. The performance data represented here were obtained using specific reagents listed in this Instruction For Use.
- All reagents should be stored refrigerated at 2-8°C in their original container. Any exceptions are clearly indicated. The reagents are stable until the expiry date when stored and handled as indicated.
- Allow all kit components and specimens to reach room temperature (22-28°C) and mix well prior to use.

- Do not interchange kit components from different lots. The expiry date printed on box and vials labels must be observed. Do not use any kit component beyond their expiry date.
- If you use automated equipment, the user has the responsibility to make sure that the kit has been appropriately tested.
- The incomplete or inaccurate liquid removal from the wells could influence the assay precision and/or increase the background. To improve the performance of the kit on automatic systems is recommended to increase the number of washes.
- It is important that the time of reaction in each well is held constant for reproducible results. Pipetting of samples should not extend beyond ten minutes to avoid assay drift. If more than 10 minutes are needed, follow the same order of dispensation. If more than one plate is used, it is recommended to repeat the dose response curve in each plate
- Addition of the TMB Substrate solution initiates a kinetic reaction, which is terminated by the addition of the Stop Solution. Therefore, the TMB Substrate and the Stop Solution should be added in the same sequence to eliminate any time deviation during the reaction.
- Observe the guidelines for performing quality control in medical laboratories by assaying control samples.
- Maximum precision is required for reconstitution and dispensation of reagents.
- Plate readers measure vertically. Do not touch the bottom of the wells.

## 6. STORAGE AND STABILITY

Store the kit at 2-8°C; the kit is stable until the expiry date claimed on the kit label and in the Certificate of Analysis.

Do not use the kit or its components after the expiry date.

## 7. PROCEDURE

### 7.1. Preparation of Calibrators and Controls

Before use, leave 5 minutes on a rotating mixer.

The Calibrators are ready to use and have the following concentration of Cortisol:

	C <sub>0</sub>	C <sub>1</sub>	C <sub>2</sub>	C <sub>3</sub>	C <sub>4</sub>
ng/mL	0	1	5	30	200

The Controls are ready to use.

Once opened, Calibrators and Controls are stable 6 months at 2-8°C.

### 7.2. Preparation of the Conjugate

The Conjugate is ready to use.

Once opened, it stable 6 months at 2-8°C.

### 7.3. Preparation of the Sample

The determination of Cortisol with this kit should be performed in urine samples.

**Important note:** the kit has been designed to be used on untreated urine samples; acidification treatments of the urine that lead the pH to values below 5.0 could interfere with the assay and produce aberrant results.

It is not necessary to dilute the sample. The total volume of urine excreted during a 24 hours should be collected and mixed in a single container.

Urine samples which are not to be assayed immediately should be stored at 2-8°C or at -20°C for longer period (maximum 6 months).

Samples with concentration greater than 200 ng/mL has not to be diluted; such samples has to be reported as "> 200 ng/mL".

#### 7.4. Preparation of Wash Solution

Dilute the content of each vial of the "10X Conc. Wash Solution" with distilled water to a final volume of 500 mL prior to use. For smaller volumes respect the 1:10 dilution ratio. The diluted wash solution is stable for 30 days at 2-8°C.

In concentrated wash solution is possible to observe the presence of crystals; in this case mix at room temperature until the complete dissolution of crystals; for greater accuracy, dilute the whole bottle of concentrated wash solution to 500 mL, taking care to transfer completely the crystals, then mix until crystals are completely dissolved.

#### 7.5. Procedure

- **Allow all reagents to reach room temperature (22-28°C) for at least 30 minutes.** At the end of the assay, store immediately the reagents at 2-8°C: avoid long exposure to room temperature.
- Unused coated microwell strips should be released securely in the foil pouch containing desiccant and stored at 2-8°C.
- To avoid potential microbial and/or chemical contamination, unused reagents should never be transferred into the original vials.
- As it is necessary to perform the determination in duplicate in order to improve accuracy of the test results, prepare two wells for each point of the calibration curve (C<sub>0</sub>-C<sub>4</sub>), two for each Control, two for each sample, one for Blank.

Reagent	Calibrator	Samples/ Controls	Blank
Calibrator C <sub>0</sub> -C <sub>4</sub>	10 µL		
Samples/ Controls		10 µL	
Conjugate	300 µL	300 µL	
Incubate at 37°C for 1 hour. Remove the contents from each well. Wash the wells 3 times with 350 µL of diluted wash solution. <b>Important note:</b> during each washing step, gently shake the plate for 5 seconds and remove excess solution by tapping the inverted plate on an absorbent paper towel. <b>Automatic washer:</b> in case you use an automatic washer, it is advised to do 6 washing steps.			
TMB Substrate	100 µL	100 µL	100 µL
Incubate at room temperature (22±28°C) for 15 minutes in the dark.			

Stop Solution	100 µL	100 µL	100 µL
Shake the microplate gently. Read the absorbance (E) at 450 nm against a reference wavelength of 620-630 nm or against Blank within 5 minutes.			

#### 8. QUALITY CONTROL

Each laboratory should assay controls at normal, high and low levels range of urinary Cortisol for monitoring assay performance. These controls should be treated as unknowns and values determined in every test procedure performed. Quality control charts should be maintained to follow the performance of the supplied reagents. Pertinent statistical methods should be employed to ascertain trends. The individual laboratory should set acceptable assay performance limits. Other parameters that should be monitored include the 80, 50 and 20% intercepts of the calibration curve for run-to-run reproducibility. In addition, maximum absorbance should be consistent with past experience. Significant deviation from established performance can indicate unnoticed change in experimental conditions or degradation of kit reagents. Fresh reagents should be used to determine the reason for the variations.

#### 9. RESULTS

##### 9.1. Mean Absorbance

Calculate the mean of the absorbance (E<sub>m</sub>) for each point of the calibration curve (C<sub>0</sub>-C<sub>4</sub>) and of each sample.

##### 9.2. Calibration curve

Plot the values of absorbance (E<sub>m</sub>) of the Calibrators (C<sub>0</sub>-C<sub>4</sub>) against concentration. Draw the best-fit curve

through the plotted points (es: Four Parameter Logistic).

### 9.3. Calculation of Results

Interpolate the values of the samples on the calibration curve to obtain the corresponding values of the concentrations expressed in ng/mL.

**To calculate the cortisol concentration in urine, calculate as above and correct for total volume of volume of urine collected in 24 hours:**

$$\text{ng/mL} \times \text{Vol(mL) urine 24 h} / 1000 = \mu\text{g Cortisol/24h}$$

## 10. REFERENCE VALUES

To determine the normal range for urine samples, 128 apparently healthy male and female adults were tested.

Result:

Normal range urine (24h)
1,5 - 63 $\mu\text{g}/24\text{h}$

## 11. PERFORMANCE AND CHARACTERISTICS

### 10.1. Analytical sensitivity

Analytical sensitivity was investigated through the LOB (white limit), the LOD (detection limit), the LOQ (quantification limit) and the anal sensitivity (A.S.).

The following table shows the criteria of the study and the results obtained.

	Criteria	Results (ng/mL)
LOB	60 replicates of Cal 0, used as "Blank", have been investigated in 5 different sessions over 3 days	0,28
LOD	6 urine samples with low cortisol concentration have been investigate over 10 assays in duplicate, performed in 5 days	0,47
LOQ	6 urine samples with low cortisol concentration have be en investigate over 10 assays in duplicate, performed in 5 days	0,56
A.S.	20 replicates of Cal 0 and 5 replicates of Cal 1 have been assayed. A.S. has been calculated by linear regression	0,22

### 10.2. Precision and reproducibility (complex precision)

Precision and reproducibility have been assessed through 6 different urine samples with different concentration of Cortisol.

The table below shows the Within Run and Total CV%.

Sample	n°	Mean (ng/mL)	Within Run CV%	Total CV%
PS2	20	112,141	6,6%	12%
PS4	20	64,563	8,1%	12%

CT High	20	50,577	7,3%	11%
PS5	20	25,878	7,6%	10%
PS6	20	9,269	7,6%	11%
CT Low	20	3,438	7,0%	9%

### 10.3. Analytical specificity

#### 10.3.1. Interfering substances

Interference for Albumin, Acetylsalicylic Acid, Ibuprofen and Ascorbic Acid were studied by adding the interfering substance to the urine sample with a low and high Cortisol concentration, and by comparing its concentration to the unspiked sample.

The interference has been evaluated as "significant" if it causes a concentration bias >10% between spiked and unspiked sample.

The following table shows the results obtained:

Substance	Concentration	Interference
Albumin	5 mg/dL	No
Acetylsalicylic acid	3,62 mmol/L	No
Ibuprofen	2,42 mmol/L	No
Ascorbic Acid	5 mg/L	No

Conclusion: no interference has been found for Albumin, Acetylsalicylic Acid, Ibuprofen and Ascorbic Acid.

#### 10.3.2. Cross-reactivity

The cross reactions of the antibody calculated at 50% according to Abraham are shown in the table:

Reagent	Cross-reactivity
Cortisol	100 %
Prednisolone	46.2 %
11-Deoxycortisol	4 %
Cortisone	3.69 %
Prednisone	3.10 %
11 $\alpha$ OH Progesterone	1 %
Progesterone	< 0.1 %
Aldosterone	< 0.1 %
Pregnenolone	< 0.1 %
17 $\beta$ Estradiol	< 0.1 %
Estrone 3-solfato	< 0.1 %
Estriol	< 0.1 %
Testosterone	< 0.1 %
Spirolactone	< 0.1 %
DHEA	< 0.1 %
DHEA-S	< 0.1 %
Androstenedione	< 0.1 %

Androsterone	< 0.1 %
DHT	< 0.1 %
Danazol	< 0.1 %
Cholesterol	< 0.1 %
Dexamethasone	< 0.1 %

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### 10.4. Correlation

137 urine samples were tested with the Urinary Cortisol Elisa kit and with a LC-MS method (reference)

The linear regression curve is:

$$Y = 1,008X - 0,5019$$

$$r^2 = 0,83$$

### 11. WASTE MANAGEMENT

Reagents must be disposed off in accordance with local regulations.

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### BIBLIOGRAPHY

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	<b>DIA.METRA SRL</b>	
Mod. PIS	<b>PACKAGING INFORMATION SHEET</b>	

**IT**  
Spiegazione dei simboli












**GB**  
Explanation of symbols

**FR**  
Explication des symboles

**ES**  
Significado de los simbolos

**DE**  
Verwendete Symbole

**PT**  
Explicação dos simbolos

	<p>DE In vitro Diagnostikum  ES Producto sanitario para diagnóstico In vitro  FR Dispositif medical de diagnostic in vitro  GB In vitro Diagnostic Medical Device  IT Dispositivo medico-diagnostico in vitro  PT Dispositivos medicos de diagnostico in vitro</p>		<p>DE Hergestellt von  ES Elaborado por  FR Fabriqué par  GB Manufacturer  IT Produttore  PT Produzido por</p>
<b>REF</b>	<p>DE Bestellnummer  ES Número de catálogo  FR Références du catalogue  GB Catalogue number  IT Numero di Catalogo  PT Número do catálogo</p>	 yyyy-mm	<p>DE Herstellungs datum  ES Fecha de fabricacion  FR Date de fabrication  GB Date of manufacture  IT Data di produzione  PT Data de produção</p>
 yyyy-mm-dd	<p>DE Verwendbar bis  ES Establa hasta (usar antes de último día del mes)  FR Utiliser avant (dernier jour du mois indiqué)  GB Use by (last day of the month)  IT Utilizzare prima del (ultimo giorno del mese)  PT Utilizar (antes ultimo dia do mês)</p>		<p>DE Biogefährdung  ES Riesco biológico  FR Risque biologique  GB Biological risk  IT Rischio biologico  PT Risco biológico</p>
 	<p>DE Gebrauchsanweisung beachten  ES Consultar las instrucciones  FR Consulter le mode d'emploi  GB Consult instructions for use  IT Consultare le istruzioni per l'uso  PT Consultar instruções para uso</p>		<p>DE Chargenbezeichnung  ES Codigo de lote  FR Numero de lot  GB Batch code  IT Codice del lotto  PT Codigo do lote</p>
 $\Sigma = xx$	<p>DE Ausreichend für "n" Tests  ES Contenido suficiente para "n" tests  FR Contenu suffisant pour "n" tests  GB Contains sufficient for "n" tests  IT Contenuto sufficiente per "n" saggi  PT Contém o suficiente para "n" testes</p>		<p>DE Inhalt  ES Contenido del estuche  FR Contenu du coffret  GB Contents of kit  IT Contenuto del kit  PT Conteúdo do kit</p>
 Max Min	<p>DE Temperaturbereich  ES Límitación de temperatura  FR Limites de température de conservation  GB Temperature limitation  IT Limiti di temperatura  PT Temperaturas limites de conservação</p>		

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## **SUGGERIMENTI PER LA RISOLUZIONE DEI PROBLEMI/TROUBLESHOOTING**

### **ERRORE CAUSE POSSIBILI/ SUGGERIMENTI**

#### **Nessuna reazione colorimetrica del saggio**

- mancata dispensazione del coniugato
- contaminazione del coniugato e/o del Substrato
- errori nell'esecuzione del saggio (es. Dispensazione accidentale dei reagenti in sequenza errata o provenienti da flaconi sbagliati, etc.)

#### **Reazione troppo blanda (OD troppo basse)**

- coniugato non idoneo (es. non proveniente dal kit originale)
- tempo di incubazione troppo breve, temperatura di incubazione troppa bassa

#### **Reazione troppo intensa (OD troppo alte)**

- coniugato non idoneo (es. non proveniente dal kit originale)
- tempo di incubazione troppo lungo, temperatura di incubazione troppa alta
- qualità scadente dell'acqua usata per la soluzione di lavaggio (basso grado di deionizzazione,)
- lavaggi insufficienti (coniugato non completamente rimosso)

#### **Valori inspiegabilmente fuori scala**

- contaminazione di pipette, puntali o contenitori- lavaggi insufficienti (coniugato non completamente rimosso)

#### **CV% intra-assy elevato**

- reagenti e/o strip non portate a temperatura ambiente prima dell'uso
- il lavatore per micropiastre non lava correttamente (suggerimento: pulire la testa del lavatore)

#### **CV% intersaggio elevato**

- condizioni di incubazione non costanti (tempo o temperatura)
- controlli e campioni non dispensati allo stesso tempo (con gli stessi intervalli) (controllare la sequenza di dispensazione)
- variabilità intrinseca degli operatori

## **ERROR POSSIBLE CAUSES / SUGGESTIONS**

### **No colorimetric reaction**

- no conjugate pipetted reaction after addition
- contamination of conjugates and/or of substrate
- errors in performing the assay procedure (e.g. accidental pipetting of reagents in a wrong sequence or from the wrong vial, etc.)

### **Too low reaction (too low ODs)**

- incorrect conjugate (e.g. not from original kit)
- incubation time too short, incubation temperature too low

### **Too high reaction (too high ODs)**

- incorrect conjugate (e.g. not from original kit)
- incubation time too long, incubation temperature too high
- water quality for wash buffer insufficient (low grade of deionization)
- insufficient washing (conjugates not properly removed)

### **Unexplainable outliers**

- contamination of pipettes, tips or containers
- insufficient washing (conjugates not properly removed) too high within-run
- reagents and/or strips not pre-warmed to CV% Room Temperature prior to use
- plate washer is not washing correctly (suggestion: clean washer head)
- too high between-run - incubation conditions not constant (time, CV % temperature)
- controls and samples not dispensed at the same time (with the same intervals) (check pipetting order)
- person-related variation