Pregnenolone ELISA
Assay Kit

Catalog Number:
PGN31-K01 (1 x 96 wells)
For Research Use Only
v. 8.0 (03.28.23)

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INTENDED USE

The Eagle Biosciences Pregnenolone ELISA Assay Kit (enzyme-linked immunoassay kit) is intended for the direct quantitative determination of Pregnenolone in human serum. The Eagle Biosciences Pregnenolone ELISA Assay Kit is for research use only and not to be used in diagnostic procedures.

LIMITATIONS RELATED TO INTENDED PURPOSE & USE

1. This test is not intended to be used for screening purposes.
2. This test is not intended for home testing or self-testing
3. The kit is calibrated for the determination of pregnenolone in human serum. The kit is not calibrated for the determination of pregnenolone in other specimens of human or animal origin.
4. The results obtained with this kit shall not be used for a clinical diagnosis and for therapeutic decisions.
5. Although common interfering substances have been evaluated with this test, other substances that have not been evaluated such as drugs and the occurrence of heterophilic antibodies in individuals regularly exposed to animals or animal products have the potential of causing interferences.

INTRODUCTION

Pregnenolone (3β-hydroxypregn-5-en-20-one) is the first steroid to be derived from cholesterol in the pathway of steroidogenesis, and it is the common precursor for all of the adrenal and gonadal steroids. Its production occurs in the mitochondrion by cleavage of the C-20 side chain of cholesterol by the P-450SCC enzyme. Once produced, pregnenolone may be utilized by two pathways of steroidogenesis. Pregnenolone may either be converted to 17-OH pregnenolone via the enzymatic action of 17α-hydroxylase or to pregnenolone via the enzymatic action of 3βhydroxysteroid dehydrogenase. Elevated pregnenolone levels occur in forms of congenital adrenal hyperplasia (CAH), due to 3β-hydroxysteroid dehydrogenase deficiency or 17αhydroxylase deficiencies. Higher levels have also been reported in women with idiopathic hirsutism. Studies on pregnenolone levels in regard to sex and age differences indicate that maximum levels occur at approximately 17 and 16 years of age for women and men, while minimum levels occur at approximately 37 and 38 years of age for women and men, respectively. In general, women were found to have slightly higher values when compared to men. Many areas of pregnenolone physiology remain to be investigated. Current research indicates that the determination of pregnenolone in serum may be useful for studying its metabolite, pregnenolone sulfate, which has been reported to have various effects in the mammalian brain and central nervous system.

PRINCIPLE OF THE ASSAY

The Pregnenolone ELISA is a competitive immunoassay. Competition occurs between pregnenolone present in calibrators, controls and specimen samples and an enzyme-labelled antigen (HRP conjugate) for a limited number of antibody binding sites on the microplate. The washing and decanting procedures remove unbound materials. After a washing step the removes unbound materials, the TMB substrate (the enzyme substrate) is added which reacts with HRP to form a blue-colored product that is inversely proportional to the amount of pregnenolone present. Following an incubation, the enzymatic reaction is terminated by addition of the stopping solution, converting the color form blue to yellow. The absorbance is measured on a microtiter plate reader at 450 nm. A set of calibrators is used to plot a calibrator curve from which the amount of pregnenolone in specimen samples and controls can be directly read.

PROCEDURAL CAUTIONS AND WARNINGS

1. This kit is for use by trained laboratory personnel (professional use only). For laboratory *in vitro* use only.
2. Practice good laboratory practices when handling kit reagents and specimens. This includes:
    a Do not pipette by mouth.
    b Do not smoke, drink, or eat in areas where specimens or kit reagents are handled.
    c Wear protective clothing and disposable gloves.
    d Wash hands thoroughly after performing the test.
    e Avoid contact with eyes, use safety glasses; in case of contact with eyes, flush eyes with water immediately and contact a doctor.
3. Users should have a thorough understanding of this protocol for the successful use of this kit. Reliable performance will only be attained by strict and careful adherence to the instructions provided.
4. Do not use this kit beyond the expiry date stated on the label.
5. If the kit reagents are visibly damaged, do not use the test kit.
6. Do not use kit components from different kit lots within a test and do not use any component beyond the expiration date printed on the label.
7. All kit reagents and specimens must be brought to room temperature and mixed gently but thoroughly before use. Avoid repeated freezing and thawing of specimens.
8. When the use of water is specified for dilution or reconstitution, use deionized or distilled water.
9. Immediately after use, each individual component of the kit must be returned to the recommended storage temperature stated on the label.
10. A calibrator curve must be established for every run.
11. It is recommended to all customers to prepare their own control materials or saliva pools which should be included in every run at a high and low level for assessing the reliability of results.
12. The controls (included in kit) must be included in every run and their results must fall within the ranges stated in the quality control certificate; a failed control result might indicate improper reagent storage.
13. When dispensing the substrate and stopping solutions, do not use pipettes in which these liquids will come into contact with any metal parts.
14. The TMB Substrate is sensitive to light and should remain colorless if properly stored. Instability or contamination may be indicated by the development of a blue color, in which case it should not be used.
15. Do not use blood contaminated saliva samples.
16. Samples or controls containing azide or thimerosal are not compatible with this kit, they may lead to false results.
17. Samples values above the measuring range of the kit may be reported as >25.6 ng/mL. If further dilution and retesting is required, only Calibrator A may be used to dilute saliva samples. The use of any other reagent may lead to false results.
18. Avoid microbial contamination of reagents.
19. To prevent the contamination of reagents, use a new disposable pipette tip for dispensing each reagent, sample, calibrator, and control.

20. To prevent contamination of reagents, do not pour reagents back into the original containers.
21. Kit reagents must be regarded as hazardous waste and disposed of according to local and/or national regulations.
22. Consumables used with the kit that are potentially biohazardous (e.g., pipette tips, bottles or containers containing human materials) must be handled according to biosafety practices to minimize the risk of infection and disposed of according to local and/or national regulations relating to biohazardous waste.
23. This kit contains 1 M sulfuric acid in the stopping solution component. Do not combine acid with waste material containing sodium azide or sodium hypochlorite.
24. The use of safety glasses, and disposable plastic, is strongly recommended when manipulating biohazardous or bio-contaminated solutions.
25. Proper calibration of the equipment used with the test, such as the pipettes and absorbance microplate reader, is required.
26. If a microplate shaker is required for the assay procedure, the type and speed of shaker required is stated in the REAGENTS AND EQUIPMENT NEEDED BUT NOT PROVIDED section. Both the type and speed of saker used can influence the optical densities and test results. If a different type of shake and/or speed is used, the user is responsible for validating the performance of the kit.
27. Do not reuse the microplate wells, they are for SINGLE USE only.
28. To avoid condensation within the microplate wells in humid environments, do not open the pouch containing the microplate until it has reached room temperature.
29. When reading the microplate, the presence of bubbles in the wells will affect the optical densities (ODs). Carefully remove any bubbles before performing the reading step.

SAFETY CAUTIONS AND WARNINGS

BIOHAZARDS

The reagents should be considered a potential biohazard and handled with the same precautions applied to blood specimens. All human specimens should be considered a potential biohazard and handled as if capable of transmitting infections and in accordance with good laboratory practices.

CHEMICAL HAZARDS

Avoid direct contact with any of the kit reagents. Specifically avoid contact with the TMB Substrate (contains tetramethylbenzidine) and Stopping Solution (contains sulfuric acid). If contacted with any of these reagents, wash with plenty of water and refer to SDS for additional information.

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SPECIMEN COLLECTION, STORAGE & PRE-TREATMENT

Specimen Collection & Storage

Approximately 0.2 mL of serum is required per duplicate determination. Collect 4–5 mL of venous blood into an appropriately labelled tube and allow it to clot. Centrifuge at room temperature and carefully transfer the serum into a new storage tube or container. Serum samples may be stored at 2-8°C for up to 24 hours or at -10°C or lower if the analyses are to be done at a later date.

Consider all human specimens as possible biohazardous materials and take appropriate precautions when handling.

Specimen Pre-Treatment

Specimen pre-treatment is not required.

REAGENTS AND EQUIPMENT NEEDED BUT NOT PROVIDED

1. Calibrated single-channel pipette to dispense 50 µL.
2. Calibrated multi-channel pipette to dispense 50 µL, 100 µL, and 150 µL.
3. Calibrated multi-channel pipettes to dispense 300 µL (if washing manually).
4. Automatic microplate washer (recommended).
5. Microplate shaker:
    a. Orbital shaker (3 mm diameter) set to 600 rpm or
    b. Reciprocating shaker (1.5" stroke length) set to 180 oscillations/minute.
6. Disposable pipette tips.
7. Distilled or deionized water.
8. Calibrated absorbance microplate reader with a 450 nm filter and an upper OD limit of 3.0 or greater.

REAGENTS PROVIDED

1. Microplate

Contents:	One anti-pregnenolone polyclonal antibody-coated 96-well (12x8) microplate in a resealable pouch with desiccant.
Format:	Ready to Use
Storage:	2-8°C
Stability:	Unopened: Stable until the expiry date printed on the label. After Opening: Stable for five weeks.

2. HRP Conjugate

Contents:	One bottle containing Pregnenolone-Horse Radish Peroxidase (HRP) conjugate in a protein-based buffer with a non-mercury preservative
Format:	Concentrated; Requires Preparation
Volume:	0.3 mL/bottle
Storage:	2-8°C
Stability:	Unopened: Stable until the expiry date printed on the label. After Opening: Stable for five weeks.
Preparation of HRP Conjugate Working Solution:	**Dilute 1:51** in assay buffer before use (e.g., 40µL of conjugate concentration in 2 mL of assay buffer). If the whole plate is to be used dilute 240 µL of conjugate concentrate in 12 mL of assay buffer. Discard any that is left over.

3. Calibrator A - F

Contents:	Six bottles of calibrator containing specified pregnenolone concentrations. Protein-based buffer with a non-mercury preservative. Prepared by spiking buffer with define quantities of pregnenolone. Listed below are approximate concentrations, please refer to vial labels for exact concentrations. Concentrations: 0, 0.1, 0.4, 1.6, 6.4, 25.6, ng/mL
Format:	Ready to Use
Volume:	Calibrator A: 2.0 mL/bottle Calibrator B-F: 0.5 mL/bottle
Storage:	2-8°C
Stability:	Unopened: Stable until the expiry date printed on the label. After Opening: Stable for five weeks.

4. Control 1 -2

Contents:	Two bottles of control containing different pregnenolone concentrations. Human serum-based buffer with a non-mercury preservative. Prepared by spiking buffer with defined quantities of pregnenolone. Refer to the QC certificate for the target values and acceptable ranges.
Format:	Ready to Use
Volume:	0.5 mL/bottle
Storage:	2-8°C
Stability:	Unopened: Stable until the expiry date printed on the label. After Opening: Stable for four weeks.

5. Assay Buffer

Contents:	One bottle containing a protein-based buffer with a non-mercury preservative
Format:	Ready to Use
Volume:	15 mL/bottle
Storage:	2-8°C
Stability:	Unopened: Stable until the expiry date printed on the label. After Opening: Stable for five weeks.

6. TMB Substrate

Contents:	One bottle containing tetramethylbenzidine and hydrogen peroxide in a non-DMF or DMSO containing buffer.
Format:	Ready to Use
Volume:	16 mL/bottle
Storage:	2-8°C

Stability:	Unopened: Stable until the expiry date printed on the label. After Opening: Stable for five weeks.

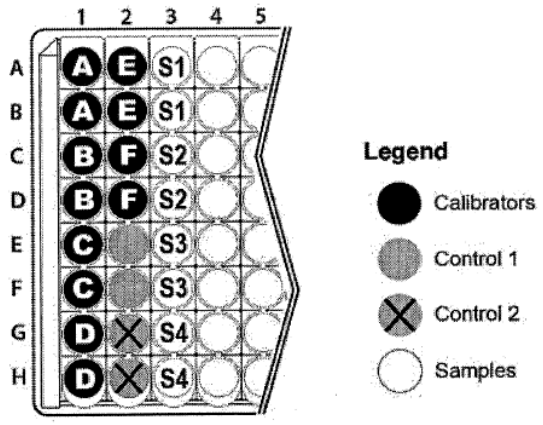
7. Stopping Solution

Contents:	One bottle containing 1M sulfuric acid.
Format:	Ready to Use
Volume:	6 mL/bottle
Storage:	2-8°C
Stability:	Unopened: Stable until the expiry date printed on the label. After Opening: Stable for five weeks.
Safety:	Refer to product SDS.

8. Wash Buffer Concentrate

Contents:	One bottle containing buffer with a non-ionic detergent and a non-mercury preservative.
Format:	Concentrated; Requires Preparation
Volume:	50 mL/bottle
Storage:	2-8°C
Stability:	Unopened: Stable until expiry date printed on the label. After Opening: Stable for five weeks. Following Preparation: The wash buffer working solution is stable for 2 weeks following preparation, assuming Good Laboratory Practices are adhered to. To prevent microbial growth, prepare the wash buffer working solution in a clean container and store under refrigerated conditions (2-8°C) when not in use.
Preparation of Wash Buffer Working Solution:	**Dilute 1:10** in distilled or deionized water before use. If the whole microplate is to be used dilute 50 mL of the wash buffer concentrate in 450 mL of distilled or deionized water.

RECOMMENDED ASSAY LAYOUT



ASSAY PROCEDURE

Specimen Pretreatment: None

All kit components, controls, and specimen samples must reach room temperature prior to use. Calibrators, controls, and specimen samples should be assayed in duplicate. Once the procedure has been started, all steps should be completed without interruption.

1. After all kit components have reached room temperature, **mix** gently by inversion.
2. **Prepare** the HRP Conjugate Working Solution and Wash Buffer Working Solution (See section *Reagents Provided, HRP Conjugate Concentration, Wash Buffer Concentrate*).
3. **Plan** the microplate wells to be used for calibrators, controls, and samples. See *Recommended Assay Layout.* Remove the strips from the microplate frame that will not be used and place them in the bag with desiccant. Reseal the bag with the unused strips and return it to the refrigerator.
4. **Pipette 50 µL** of each calibrator, control, and pre-treated specimen sample into assigned wells.
5. **Pipette 100 µL** of the HRP conjugate Working Solution into each well (the use of a multi-channel pipette is recommended).
6. **Incubate** the microplate on a microplate shaker** for **60 minutes** at room temperature.
7. **Wash** the microplate wells with an automatic microplate washer (preferred) or manually as state below.
    a **Automatic:** Using an automatic microplate washer, perform a **3-cycle** wash using **300 µL /well** of Wash Buffer Working Solution (3 x 300 µL). One cycle consists of aspirating all wells then filling each well with 300 µL of Wash Buffer Working Solution. After the final wash cycle, aspirate all wells and then tap the microplate firmly against absorbent paper to remove any residual liquid.
    b **Manually:** For manual washing, perform a **3-cycle** wash using **300 µL /well** of Wash Buffer Working Solution (3 x 300 µL). One cycle consists of aspirating all wells by briskly emptying the contents of the wells over a waster container, then pipetting 300 µL of Wash Buffer Working Solution into each well using a multi-channel pipette. After the final wash cycle, aspirate all wells by briskly emptying the contents over a waste container and then tap the microplate firmly against absorbent paper to remove any residual liquid.
8. **Pipette 150 µL** of TMB Substrate into each well (the use of a multi-channel pipette is recommended).
9. **Incubate** the microplate on a microplate shaker** for **10-15 minutes** at room temperature.
10. **Pipette 50 µL** of Stopping Solution into each well (the use of a multi-channel pipette is recommended) in the same order and speed as was used for the addition of TMB Substrate. Gently tap the microplate frame to mix the contents of the wells.
11. **Measure** the optical density (absorbance) in the microplate wells using an absorbance microplate reader set to 450 nm, within 20 minutes after addition of the Stopping Solution.

**See Section *Reagents and Equipment Needed But Not Provided* for microplate shaker options

CALCULATIONS

1. Calculate the mean optical density of each calibrator, control, and specimen sample duplicate.
2. Use a 4-parameter or 5-parameter curve fit with immunoassay software to generate a calibrator curve.
3. The immunoassay software will calculate the concentrations of the controls and specimen samples using the mean optical density values and the calibrator curve.
4. If a sample reads more than 25.6 ng/mL and needs to be diluted and retested, then dilute it with calibrator A not more than 1:8. The result obtained should be multiplied by the dilution factor.

QUALITY CONTROL

When assessing the validity of the test results, the following criteria should be evaluated:

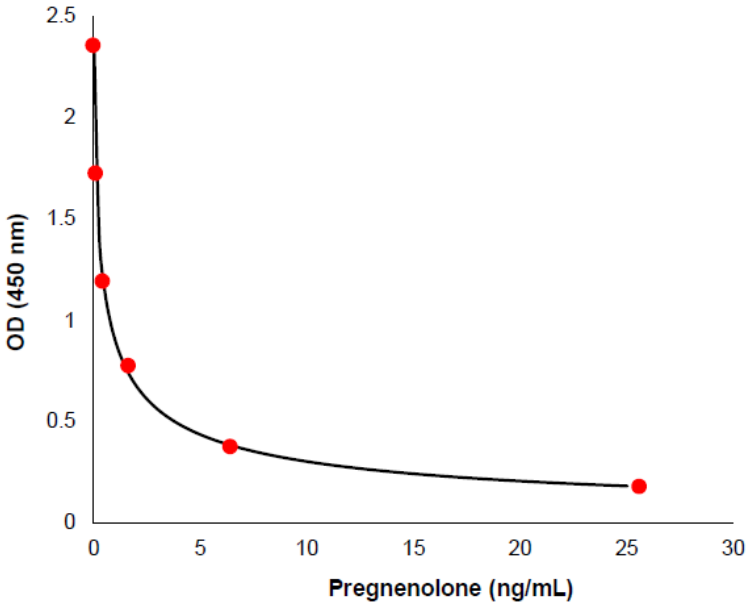
1. The Calibrator A mean optical density meets the acceptable range as stated in the QC Certificate
2. The calibrator with the highest concentration meets the % binding acceptable range as stated in the QC Certificate. % Binding = (OD of calibrator / OD of calibrator A) X 100.
3. The values obtained for the kit controls are within the acceptable ranges as stated in the QC Certificate.
4. The results of any external controls that were used meet the acceptable ranges.

TYPICAL TABULATED DATA

Sample data only. Do not use to calculate results.

Calibrator	Mean OD (450 nm)	% Binding	Value (ng/mL)
A	2.357	100	0
B	1.722	73	0.1
C	1.190	50	0.4
D	0.744	33	1.6
E	0.377	16	6.4
F	0.177	7	25.6
Unknown	0.480	-	4.1

TYPICAL CALIBRATOR CURVE



PERFORMANCE CHARACTERISTICS SENSITIVITY

The lower detection limit is calculated from the standard curve by determining the resulting concentration of the mean OD of Calibrator A (based on 10 replicate analyses) minus 2 SD. Therefore, the sensitivity of the Pregnenolone ELISA kit is 0.05 ng/mL.

SPECIFICITY (CROSS-REACTIVITY)

The following compounds were tested for cross-reactivity with the Pregnenolone ELISA kit with pregnenolone cross-reacting at 100%.

Steroid	% Cross Reactivity
Pregnenolone	100
Pregnenolone	6.0
Dehydroisoandrosterone	5.2
5a-Androstandiol	4.7
Epiandrosterone	1.0
Pregnenolone Sulfate	0.4
Androstandione	0.3
5a-Androsterone	0.3
DHEAS	0.2
Etiocholanolone	0.1

The following steroids were tested but cross-reacted at less than 0.1%: Androsterone, Aldosterone, Androstenedione, Cholesterol, Corticosterone, 5α-DHT, 17β-Estradiol, Estriol and Testosterone.

INTRA-ASSAY PRECISION

Three samples were assayed ten times each on the same calibrator curve. The results (in ng/mL) are tabulated below:

Sample	Mean	SD	CV %
1	0.19	0.02	10.6
2	1.04	0.85	8.2
3	4.77	0.37	7.8

INTER-ASSAY PRECISION

Three samples were assayed ten times over a period of four weeks. The results (in ng/mL) are tabulated below:

Sample	Mean	SD	CV %
1	0.22	0.03	14.5
2	1.14	0.14	12.3
3	4.56	0.44	9.6

RECOVERY

Spike samples were prepared by adding defined amounts of pregnenolone to four serum samples, The results (in ng/mL) are tabulated below:

Sample	Obs. Result	Exp. Result	Recovery %
1 Unspiked +4.14	0.37 5.31	- 4.51	- 117.7
2 Unspiked +4.01	0.77 5.69	- 4.78	- 119.0
3 Unspiked +3.98	0.85 5.18	- 4.83	- 107.2
4 Unspiked +3.78	1.47 6.31	- 5.25	- 120.2

LINEARITY

Three patient serum samples were diluted with calibrator A. The results (in ng/mL) are tabulated below:

Sample	Obs. Result	Exp. Result	Recovery %
1 1:2 1:4 1:8	5.31 2.89 1.26 0.71	- 2.66 1.33 0.66	- 108.6 94.7 107.6
2 1:2 1:4 1:8	6.51 2.75 1.54 0.80	- 3.26 1.63 0.81	- 84.4 94.5 98.8
3 1:2 1:4 1:8	8.34 3.78 2.15 1.05	- 4.17 2.09 1.04	- 90.6 102.9 101.0

REFERENCE RANGES

As for all clinical assays each laboratory should collect data and establish their own range of expected normal values.

Group	N	Mean (ng/mL)	Abs. Range (ng/mL)
Males	30	0.50	0.1-3.4
Females	50	0.55	0.1-3.8

REFERENCES

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WARRANTY INFORMATION

Eagle Biosciences, Inc. warrants its Product(s) to operate or perform substantially in conformance with its specifications, as set forth in the accompanying package insert. This warranty is expressly limited to the refund of the price of any defective Product or the replacement of any defective Product with new Product. This warranty applies only when the Buyer gives written notice to the Eagle Biosciences within the expiration period of the Product(s) by the Buyer. In addition, Eagle Biosciences has no obligation to replace Product(s) as result of a) Buyer negligence, fault, or misuse, b) improper use, c) improper storage and handling, d) intentional damage, or e) event of force majeure, acts of God, or accident.

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For further information about this kit, its application or the procedures in this kit, please contact the Technical Service Team at Eagle Biosciences, Inc. at info@eaglebio.com or at 866-411-8023.