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Human Total FGF-21 ELISA Assay Kit

Catalog Number:

T2131-K01 (1 x 96 wells)

For Research Use Only. Not for use in diagnostic procedures.

v. 1.1 (09.19.18)

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I. INTENDED USE

The Eagle Biosciences Total FGF-21 ELISA Assay Kit is intended for the quantitative determination of human N-terminal and C-terminal FGF-21 level in EDTA-plasma or serum. It measures the intact FGF-21, the N-terminal and C-terminal FGF-21 fragments that must not be N-terminally and C-terminally truncated. The Eagle Biosciences Total FGF-21 ELISA Assay Kit is intended for research use only.

II. INTRODUCTION

Fibroblast Growth Factor 21 (FGF-21) belongs to the FGF-19 subfamily, which includes FGF-19, FGF-21 and FGF-23. The FGF-19 family members are potent endocrine hormones in the regulation of a diverse physiological homeostasis.

The intact FGF-21 is a small protein comprising 181 amino acids. Administration of recombinant FGF-21 lowered plasma glucose and insulin levels, reduced hepatic and circulating triglycerides and cholesterol levels, and improved insulin sensitivity, energy expenditure, hepatic steatosis and obesity in a range of insulin-resistant animal models. The physiological functions of FGF-21 are relied on the intact molecular structure and amino acid sequence in its N-terminal and C-terminal region. The C-terminal non-truncated FGF-21 is a potent cell membrane β -Klotho binder. Whereas, a C-terminal truncated FGF-21 (1-170) is a potent inhibitor that competitively inhibits the biological activity of intact FGF-21 (1-181). Therefore, it is important to measure the circulation intact FGF-21 level in the assessment of the physiological and pathophysiological condition. An assay that determines the fragment of the FGF-21 might overestimate the biological activity of the protein in test sample.

Circulation FGF-21 is a biomarker and its levels are increased in patients with nonalcoholic fatty liver disease (NAFLD), type 2 diabetes, gestational diabetes and obesity. An increase of circulating FGF-21 is also found in patients with Cushing's syndrome, patients with lipodystrophy induced by HIV-1 and patients with chronic renal disease or end-stage renal disease (ESRD).

III. ASSAY PRINCIPLE

The Eagle Biosciences Total FGF-21 ELISA Assay Kit is designed, developed and produced for the quantitative determination of human N-terminal and C-terminal FGF-21 level in EDTA-plasma or serum. It measures the intact FGF-21, the N-terminal and C-terminal FGF-21 fragments that must not be N-terminally and C-terminally truncated. The assay utilizes the two-site "sandwich" technique with two selected antibodies that bind to different epitopes of human FGF-21. One of the antibodies specifically binds to the C-terminal human FGF-21 (175-181) and the other is to the multi-epitopes of mid-regional and N-terminal human FGF-21.

Assay calibrators, controls and patient samples are added directly to wells of a microplate that is coated with an anti-human FGF-21 (175-181) specific antibody. After the first



incubation period, a horseradish peroxidase-conjugated anti-human FGF-21 polyclonal antibody is added to each well. After the second incubation period, the antibody on the wall of microtiter well captures human N-terminal and C-terminal FGF-21 in the sample and further forms "sandwich" with the tracer antibody. Unbound proteins in each microtiter well are washed away. An immunocomplex of "anti-FGF-21 antibody --- human N-terminal and C-terminal FGF-21 --- HRP-conjugated tracer antibody" is formed. The unbound tracer antibody is removed in the subsequent washing step. For the detection of this immunocomplex, the well is then incubated with a substrate solution in a timed reaction and then measured in a spectrophotometric microplate reader. The enzymatic activity of the immunocomplex bound to human N-terminal and C-terminal FGF-21 on the wall of the microtiter well is directly proportional to the amount of N-terminal and C-terminal FGF-21 in the sample. A calibrator curve is generated by plotting the absorbance versus the respective human intact FGF-21 concentration for each calibrator on point-to-point or 4 parameter curve fit. The concentration of the combined N-terminal and C-terminal FGF-21 in test samples is determined directly from this calibrator curve.

IV. REAGENTS: Preparation and Storage

The Eagle Biosciences Total FGF-21 ELISA Assay kit must be stored at 2 – 8 °C upon receipt. For the expiration date of the kit refer to the label on the kit box. All components are stable until this expiration date.

Prior to use allow all reagents to come to room temperature. Regents from different kit lot numbers should not be combined or interchanged.

1. **Anti-Human ncFGF-21 Antibody Coated Microplate**

One microplate with 12 x 8 well-breakable strips (96 wells total) coated with antibody to human N-terminal FGF-21. The plate is framed and sealed in a foil zipper bag with a desiccant. This reagent should be stored at 2 – 8°C and is stable until the expiration date on the kit box.

2. **Human ncFGF-21 Tracer Antibody**

One vial containing 0.6 mL concentrated HRP-labeled anti-human FGF-21 polyclonal antibody in a stabilized protein matrix. This reagent must be diluted with FGF-21 Tracer Antibody Diluent before use. This reagent should be stored at 2 – 8°C and is stable until the expiration date on the kit box.

3. **FGF-21 Tracer Antibody Diluent**

One vial containing 12 mL ready-to-use buffer. It should be only used for tracer antibody dilution according to the assay procedures. This reagent should be stored at 2 – 8°C and is stable until the expiration date on the kit box.

4. **ELISA Wash Concentrate**

One bottle contains 30 mL of 30-fold concentrate. Before use the contents must be diluted with 870 mL of demineralized water and mixed well. Upon dilution this



yields a working wash solution containing a surfactant in phosphate buffered saline with a non-azide, non-mercury preservative. The diluted wash solution should be stored at room temperature and is stable until the expiration date on the kit box.

5. ELISA HRP Substrate

One bottle contains 12 mL of tetramethylbenzidine (TMB) with stabilized hydrogen peroxide. This reagent should be stored at 2 – 8°C and is stable until the expiration date on the kit box.

6. ELISA Stop Solution

One bottle contains 12 mL of 0.5 M sulfuric acid. This reagent should be stored at 2 – 8°C or room temperature and is stable until the expiration date on the kit box.

7. Human FGF-21 Calibrators

Six vials each contain different concentrations of human FGF-21 in a lyophilized bovine serum-based matrix with a non-azide, non-mercury preservative. **Refer to vials for exact concentration for each calibrator.** These reagents should be stored at 2 – 8°C and are stable until the expiration date on the kit box.

8. Human FGF-21 Controls

Two vials each contain different concentrations of human FGF-21 in a lyophilized bovine serum- based matrix with a non-azide, non-mercury preservative. Refer to vials for exact concentration range for each control. Both controls should be stored at 2 – 8°C and are stable until the expiration date on the kit box.

V. SAFETY PRECAUTIONS

The reagents must be used in a professional setting by trained personnel. The source material for reagents containing bovine serum was derived in the contiguous 48 United States. It was obtained only from healthy donor animals maintained under veterinary supervision and found free of contagious diseases.

Wear gloves while performing this assay and handle these reagents as if they are potentially infectious. Avoid contact with reagents containing TMB, hydrogen peroxide, or hydrochloric acid. TMB may cause irritation to skin and mucous membranes and cause an allergic skin reaction. TMB is a suspected carcinogen. Hydrochloric acid may cause severe irritation on contact with skin. Provide good ventilation in process area to prevent formation of vapor. Do not breathe mist, vapors, spray. Do not get in eyes, on skin, or on clothing. Do not ingest or inhale fumes. On contact, flush with copious amounts of water for at least 15 minutes. Use Good Laboratory Practices.

VI. MATERIALS REQUIRED BUT NOT PROVIDED

1. Precision single channel pipettes capable of delivering 25 µL, 50 µL, 100 µL, and 1000 µL etc.



2. Repeating dispenser suitable for delivering 100 μ L.
3. Disposable pipette tips suitable for above volume dispensing.
4. Disposable 12 x 75 mm or 13 x 100 glass or plastic tubes.
5. Disposable plastic 100 mL and 1000 mL bottle with caps.
6. Aluminum foil.
7. Deionized or distilled water.
8. Plastic microtiter well cover or polyethylene film.
9. ELISA multichannel wash bottle or automatic (semi-automatic) washing system.
10. Spectrophotometric microplate reader capable of reading absorbance at 450 nm.

VII. SPECIMEN COLLECTION

Serum or EDTA plasma are acceptable samples.

Only 100 μ L of human serum or EDTA plasma sample is required for human cFGF-21 measurement in singlet. No special preparation of individual is necessary prior to specimen collection. Whole blood should be collected with lavender-top Vacutainer. Separate the plasma from cells by centrifugation (850 – 1500xg for 10 minutes). The samples should be separated from the cells right after collection or at least within one hour of blood collection and should be transferred to a clean test tube right after centrifugation. Serum and EDTA plasma samples can be stored at 2-8°C for no more than 72 hours, otherwise must be stored at -20°C. Avoid more than three freeze-thaw cycles of specimen.

Specimen Shipment

Collected EDTA-plasma or serum samples should be shipped to designated laboratory in frozen condition with dry ice. If frozen condition is not available, samples should be shipped with blue ice in an insulated container for a maximum of 48 hours.

VIII. ASSAY PREPARATION

1. Reagent Preparation

1. Prior to use allow all reagents to come to room temperature. Reagents from different kit lot numbers should not be combined or interchanged.
2. ELISA Wash Concentrate must be diluted to working solution prior to use. Please see REAGENTS section for details.
3. Reconstitute kit calibrators and controls by adding 0.5 mL distilled water into each vial. Gently mix and dissolve the entire particle before use. The reconstituted calibrators and controls should be stored at -20°C right after use.
4. Prepare working human ncFGF-21 tracer antibody by 1:21 fold dilution of the conjugation antibody with the nFGF-21 Tracer Antibody Diluent. Following is a table that outlines the relationship of strips used and antibody mix prepared.



Strip no.	nFGF-21 Tracer Antibody Diluent	nFGF-21 Tracer Antibody
1	1000 μ L	50 μ L
2	2000 μ L	100 μ L
3	3000 μ L	150 μ L
4	4000 μ L	200 μ L
5	5000 μ L	250 μ L
6	6000 μ L	300 μ L
7	7000 μ L	350 μ L
8	8000 μ L	400 μ L
9	9000 μ L	450 μ L
10	1000 μ L	500 μ L
11	1100 μ L	550 μ L
12	1200 μ L	600 μ L

Note: this antibody mixture must be freshly prepared right before testing.

VIII. ASSAY PROCEDURE

Test Configuration

Row	STRIP 1	STRIP 2	STRIP 3
A	CAL 1	CAL 5	SAMPLE 1
B	CAL 1	CAL 5	SAMPLE 1
C	CAL 2	CAL 6	SAMPLE 2
D	CAL 2	CAL 6	SAMPLE 2
E	CAL 3	CTL 1	SAMPLE 3
F	CAL 3	CTL 1	SAMPLE 3
G	CAL 4	CTL 2	
H	CAL 4	CTL 2	

1. Place a sufficient number of antibody coated microwell strips in a holder to run human FGF-21 calibrators, controls and unknown samples in duplicate.
2. Add 100 μ L of calibrators, controls and patient plasma/serum samples into the designated microwell.
3. Cover the plate with one plate sealer and incubate plate with orbital shaking 170 rpm (big radius) or 400 rpm (smaller radius) at room temperature for 1 hour.
4. Remove plate sealer. Aspirate the contents of each well. Wash each well 5 times by dispensing 350 μ L of working wash solution into each well and then completely aspirating the contents. Alternatively, an automated microplate washer can be used.
5. Add 100 μ L of 1:21 diluted tracer antibody to each well.
6. Cover the plate with one plate sealer and incubate plate with orbital shaking 170 rpm (big radius) or 400 rpm (smaller radius) at room temperature for 1 hour.
7. Remove plate sealer. Aspirate the contents of each well. Wash each well 5 times by dispensing 350 μ L of working wash solution into each well and then completely



aspirating the contents. Alternatively, an automated microplate washer can be used.

8. Add 100 μL of ELISA HRP Substrate into each of the wells.
9. Cover the plate with one plate sealer and also with aluminum foil to avoid exposure to light. Incubate plate at room temperature for 20 minutes.
10. Remove the aluminum foil and plate sealer. Add 100 μL of ELISA Stop Solution into each of the wells. Mix gently.
11. Read the absorbance at 450/650 nm within 10 minutes in a microplate reader.

X. PROCEDURAL NOTES

1. It is recommended that all calibrators, controls and unknown samples be assayed in duplicate. The average absorbance reading of each duplicate should be used for data reduction and the calculation of results.
2. Keep light-sensitive reagents in the original amber bottles.
3. Store any unused antibody-coated strips in the foil zipper bag with desiccant to protect from moisture.
4. Careful technique and use of properly calibrated pipetting devices are necessary to ensure reproducibility of the test.
5. Incubation times or temperatures other than those stated in this insert may affect the results.
6. Avoid air bubbles in the microwell as this could result in lower binding efficiency and higher CV% of duplicate reading.
7. All reagents should be mixed gently and thoroughly prior to use. Avoid foaming.

XI. INTERPRETATION OF RESULTS

1. Calculate the average absorbance for each pair of duplicate test results.
2. Subtract the average absorbance of the CAL 1 (0 pg/mL) from the average absorbance of all other readings to obtain corrected absorbance.
3. The calibrator curve is generated by the absorbance of all calibrators. Appropriate computer assisted data reduction programs may also be used for the calculation of results.
- 4.

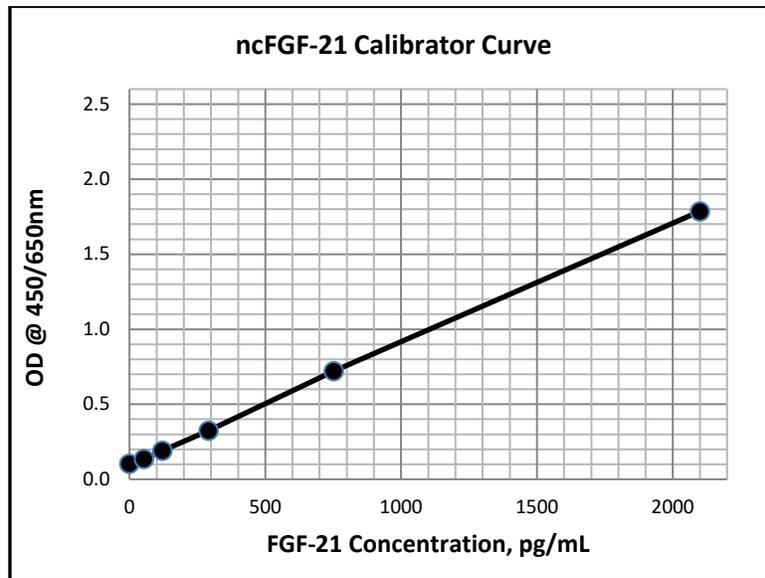
The human Total FGF-21 concentrations for the controls and patient samples are read directly from the calibrator curve using their respective corrected absorbance.

XII. EXAMPLE DATA AND STANDARD CURVE

A typical absorbance data and the resulting standard curve from human FGF-21 ELISA are represented. **This curve should not be used in lieu of standard curve generated with each assay.**



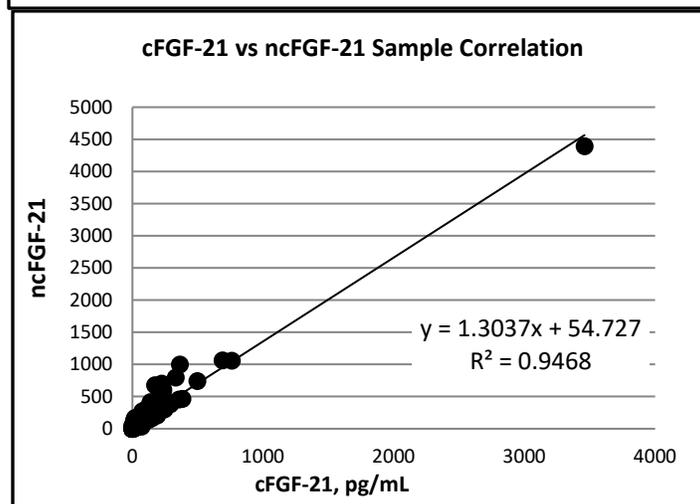
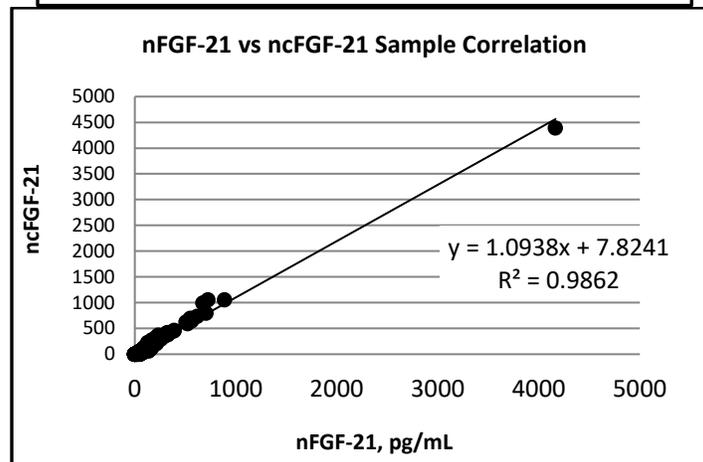
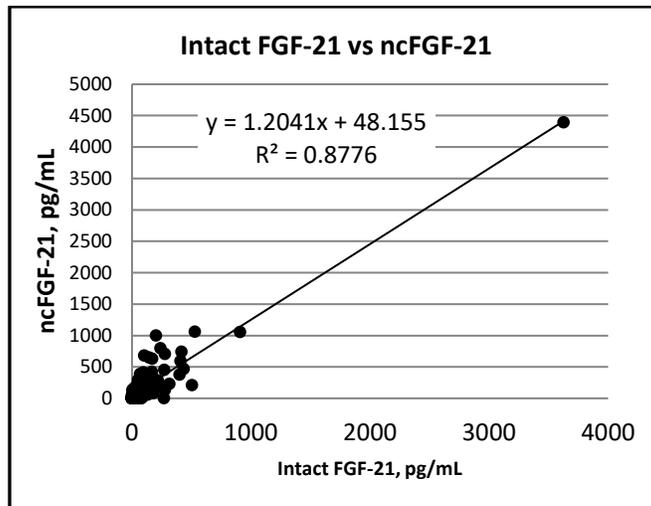
Well I.D.	OD 450/620 nm Absorbance			Results (mIU/L)
	Readings	Average	Corrected	
CAL-1: 0 pg/mL	0.103 0.105	0.104	0.000	
CAL-2: 54.0 pg/mL	0.133 0.140	0.137	0.033	
CAL-3: 121 pg/mL	0.200 0.184	0.192	0.088	
CAL-4: 291 pg/mL	0.322 0.326	0.324	0.220	
CAL-5: 752 pg/mL	0.711 0.731	0.721	0.617	
CAL-6: 2100 pg/mL	1.781 1.791	1.786	1.682	
Control 1	0.246 0.244	0.245	0.141	189.1
Control 2	0.524 0.525	0.525	0.421	523.6





XIII. EXPECTED VALUES

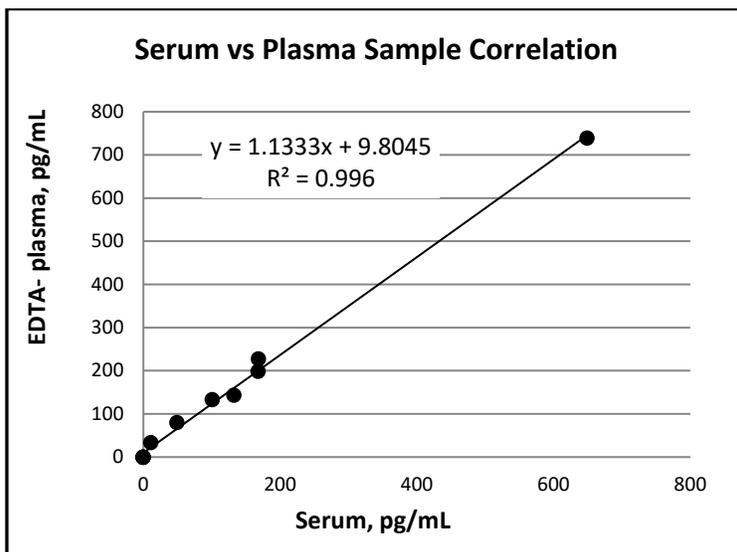
This human Total FGF-21 ELISA was validated by testing the sample correlation against human intact FGF-21 (F2131-K01), N-Terminal FGF-21 (N2131-K01), and C-Terminal FGF-21 (CTF31-K01). Total of combined 138 serum/plasma samples were measured.





The normal range was found to be less than 200 pg/mL. It is strongly recommended that each laboratory should establish its own normal range based on normal donor EDTA-plasma and serum samples

Total of 11 EDTA plasma and 16 serum samples were measured side-by-side with this ELISA kit. It was found that EDTA-plasma samples give a slightly higher value than serum samples. It is recommended to use serum samples for this assay.



XIV. LIMITATION OF THE PROCEDURE

1. Since there is no Gold Standard concentration available for human N-terminal and C-terminal FGF-21 measurement, the values of assay calibrators were established by correlation to a highly purified FGF-21 standard.
2. For sample values reading greater than the highest calibrator, it is recommended to re-assay samples with dilution.
3. Bacterial or fungal contamination of plasma specimens or reagents, or cross-contamination between reagents may cause erroneous results.
4. Water deionized with polyester resins may inactivate the horseradish peroxidase enzyme.

XV. QUALITY CONTROL

To assure the validity of the results each assay should include adequate controls with known human N-terminal and C-terminal FGF-21 levels. We recommend that all assays include the laboratory's own FGF-21 controls in addition to those provided with this kit.

XV. PERFORMANCE CHARACTERISTICS

Sensitivity

The sensitivity (lowest limit of detection) of this human nFGF-21 ELISA as determined by the corresponding OD value of 2-fold standard deviation above the mean on 16 duplicate



determination of zero calibrator is 10.4 pg/mL. The Limit of Quantitation in 95th percentile is 20.6 pg/mL.

High Dose “hook” effect

This assay has showed that it did not have any high dose “hook” effect up to 236, 900 pg/mL.

Precision

The intra-assay precision is validated by measuring three donor EDTA-plasma samples in a single assay with 12 replicate determinations.

Mean Human ncFGF-21 Value (pg/mL)	CV (%)
275.9	9.8
673.5	5.2

The inter-assay precision is validated by measuring three control samples in duplicate in 8 individual assays.

Mean Human ncFGF-21 Value (pg/mL)	CV (%)
196.8	6.4
517.2	4.3

Linearity

Linearity was validated using Level 5 and Level 6, diluted with Calibrator Matrix and assayed

	Expected	Observed	% Recovery
Level 6	-	2100	-
1:2	1050	1069.888	101.9%
1:4	525	504.475	96.1%
1:8	262.5	242.964	92.6%
Level 5	-	700	-
1:2	350	333.188	95.2%
1:4	175	160.554	91.7%
1:8	87.5	76.716	87.7%



Spike Recovery

Two serum samples were spiked, in equal volume, with various amounts of human ncFGF-21 and assayed. The results in the value of pg/mL are as follows:

	Expected	Observed	% Recovery
Sample A	-	46.769	-
+CAL 3: 77.8	62.2845	71.724	115.2%
+CAL 4: 233.3	140.0345	142.964	102.1%
+CAL 5: 700	373.3845	359.668	96.3%
Sample B	-	92.965	-
+CAL 3: 77.8	85.3825	80.456	94.2%
+CAL 4: 233.3	163.1325	147.121	90.2%
+CAL 5: 700	396.4825	359.861	90.8%

XVI. REFERENCES

1. Yie J, et al. FGF21 N- and C-termini play different roles in receptor interaction and activation. FEBS Lett. 2009 Jan 5;583:19-24.
2. Micanovic R, et al. Different roles of N- and C- termini in the functional activity of FGF21. J Cell Physiol. 2009 May;219(2):227-34.
3. Yusuke Murata, et al. FGF21 as an Endocrine Regulator in Lipid Metabolism: From Molecular Evolution to Physiology and Pathophysiology. Journal of Nutrition and Metabolism, Vol 2011, Article ID 981315, 8 pages

Warranty Information

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For further information about this kit, its application or the procedures in this kit, please contact the Technical Service Team at Eagle Biosciences, Inc. at info@eaglebio.com or at 866-411-8023.