Dengue Virus IgM ELISA Kit

Enzyme Immunoassay for the quantification of Dengue Virus IgM in serum and plasma

Catalog number: ARG80539
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**MANUFACTURED BY:**

Arigo Biolaboratories Corporation

Address: No. 22, Ln. 227, Gongyuan Rd., Hsinchu City 300, Taiwan

Phone: +886 (3) 562 1738

Fax: +886 (3) 561 3008

Email: info@arigobio.com

www.arigobio.com
INTRODUCTION

Dengue virus is a single-stranded RNA virus of about 50 nm in diameter belonging to the genus Flavivirus. Dengue and dengue hemorrhagic fever are caused by one of four closely related, but antigenically distinct, virus serotypes (DEN-1, DEN-2, DEN-3, and DEN-4). Infection with one of these serotypes does not provide cross protective immunity, so persons living in a dengue-endemic area can have four dengue infections during their lifetimes. The viruses are transmitted by Aedes aegypti, a domestic, day-biting mosquito that prefers to feed on humans. Infection with dengue viruses produces a spectrum of clinical illness ranging from a nonspecific viral syndrome to severe and fatal hemorrhagic disease. It is primarily a disease of the tropics; its global distribution is comparable to that of malaria, and an estimated 2.5 billion people live in areas at risk for epidemic transmission. Globally, there are an estimated 50 to 100 million cases of dengue fever and several hundred thousand cases of dengue hemorrhagic fever.

- The case-fatality rate of DHF in most countries is about 5%; most fatal cases are among children and young adults.
- Important risk factors for DHF include the strain and serotype of the infecting virus, as well as the age, immune status, and genetic predisposition of the patient.
- Risk groups: residents of or visitors to tropical urban areas.

<table>
<thead>
<tr>
<th>Species</th>
<th>Disease</th>
<th>Symptoms</th>
<th>Mechanism of Infection</th>
</tr>
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<tbody>
<tr>
<td>Dengue virus</td>
<td>Dengue</td>
<td>Sudden onset of fever, severe headache, myalgia and arthralgia leukopenia, thrombocytopenia and hemorrhagic manifestations</td>
<td>Transmission by mosquitos (Aedes aegypti)</td>
</tr>
<tr>
<td></td>
<td>Dengue hemorrhagic fever (DHF) or Breakbone fever</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>
The presence of virus resp. infection may be identified by

- Serology: Detection of antibodies by ELISA

Infection produces lifelong immunity, but the antigenically distinct serotypes do not provide cross-protective immunity, so a person can theoretically experience four dengue infections; a dengue vaccine is not available.

**PRINCIPLE OF THE ASSAY**

This assay employs the quantitative enzyme immunoassay technique. A specific Dengue Virus antigen Type 2 has been pre-coated onto a microtiter plate. Standards or samples are pipetted into the wells and any Dengue virus antibody present is bound by the immobilized antigen. After washing away any unbound substances, an HRP-conjugated antibody specific for human IgM is added to each well and incubate. Following the washing of any unbound antibody-enzyme reagent, a substrate solution (TMB) is added to the wells and color develops in proportion to the amount of antigen-antibody binding in the initial step. The color development is stopped by the addition of acid and the intensity of the color is measured at a wavelength of 450nm ±2nm.
MATERIALS PROVIDED & STORAGE INFORMATION

Store the unopened kit at 2-8 °C. Use the kit before expiration date.

<table>
<thead>
<tr>
<th>Component</th>
<th>Quantity</th>
<th>Storage information</th>
</tr>
</thead>
<tbody>
<tr>
<td>Antigen-coated microplate</td>
<td>8 X 12 strips</td>
<td>4°C. Unused strips should be sealed tightly in the air-tight pouch.</td>
</tr>
<tr>
<td>Calibrator A (Negative Control)</td>
<td>2ml</td>
<td>4°C</td>
</tr>
<tr>
<td>Calibrator B (Cut-off Standard)</td>
<td>3ml</td>
<td>4°C</td>
</tr>
<tr>
<td>Calibrator C (Weak Positive Control)</td>
<td>2ml</td>
<td>4°C</td>
</tr>
<tr>
<td>HRP-conjugated antibody</td>
<td>20ml (Ready-to-use)</td>
<td>4°C</td>
</tr>
<tr>
<td>Sample Diluent</td>
<td>100ml</td>
<td>4°C</td>
</tr>
<tr>
<td>20X Wash buffer</td>
<td>50ml</td>
<td>4°C</td>
</tr>
<tr>
<td>TMB substrate</td>
<td>15ml</td>
<td>4°C (Protect from light)</td>
</tr>
<tr>
<td>STOP solution</td>
<td>15ml</td>
<td>4°C</td>
</tr>
</tbody>
</table>

MATERIALS REQUIRED BUT NOT PROVIDED

- Microplate reader capable of measuring absorbance at 450nm
- Pipettes and pipette tips
- Deionized or distilled water
- Automated microplate washer (optional)

TECHNICAL HINTS AND PRECAUTIONS

- Wear protective gloves, clothing, eye, and face protection especially while handling blood or body fluid samples.
**Dengue Virus IgM ELISA Kit ARG80539**

- Store the kit at 4°C at all times.
- Briefly spin down the antibody conjugate concentrate and HRP-Streptavidin concentrate before use.
- If crystals are observed in the 20X Wash buffer, warm to RT (not more than 50°C) until the crystals are completely dissolved.
- Ensure complete reconstitution and dilution of reagents prior to use.
- It is highly recommended that the standards, samples and controls be assayed in duplicates.
- Change pipette tips between the addition of different reagent or samples.

**SAMPLE COLLECTION & STORAGE INFORMATION**

The sample collection and storage conditions listed below are intended as general guidelines. Sample stability has not been evaluated.

**Serum** - Use a serum separator tube (SST) and allow samples to clot for 30 minutes before centrifugation for 15 minutes at 1000 x g. Remove serum and assay immediately or aliquot and store samples at ≤ -20 °C. Avoid repeated freeze-thaw cycles.

**Plasma** - Collect plasma using Citrate as an anticoagulant. Centrifuge for 15 minutes at 1000 x g within 30 minutes of collection. Assay immediately or aliquot and store samples at ≤ -20 °C. Avoid repeated freeze-thaw cycles.

For the performance of the test, the samples have to be diluted 1:101 with sample diluent.
REAGENT PREPARATION

- 1X Wash buffer: Dilute 20X Wash buffer into distilled water to yield 1X Wash buffer.
- Samples: Samples have to be diluted 1:100 with sample diluent.

ASSAY PROCEDURE

All materials should be equilibrated to room temperature (RT) before use. Standards, samples and controls should be assayed in duplicates.

1. Remove excess microplate strips from the plate frame, return them to the foil pouch containing the desiccant pack, and reseal it.
2. Add 100 μl of controls, diluted samples (1:100) and zero controls (sample diluent buffer) into wells. Incubate for 1h at 37°C.
3. Aspirate each well and wash, repeating the process 4 times for a total 5 washes. Wash by filling each well with 1x Wash Buffer (300 μl) using a squirt bottle, manifold dispenser, or autowasher. Complete removal of liquid at each is essential to good performance. After the last wash, remove any remaining Wash Buffer by aspirating, decanting or blotting against clean paper towels.
4. Add 100 μl HRP-conjugated antibody (ready-to-use) into each well. Cover wells and incubate for 30 minutes at RT.
5. Aspirate each well and wash as step 3.
6. Add 100 μl of TMB Reagent to each well. Incubate for 15 minutes at room temperature.
7. Add 100 μl of Stop Solution to each well. The color of the solution should
change from blue to yellow.

8. Read the OD with a microplate reader at 450nm immediately.

**CALCULATION OF RESULTS**

1. In order for an assay to be considered valid, the following criteria must be met:
   - Substrate blank: Absorbance value <0.1
   - Negative control: Absorbance value <0.2 and <cut-off
   - Cut-off control: Absorbance value 0.15-1.3
   - Positive control: Absorbance value >cut-off
   - If these criteria are not met, the test is not valid and must be repeated.

2. Interpretation of results:
   - The samples are considered positive if the absorbance value is higher than 10% over the cut-off.
   - Samples with absorbance value of 10% above or below cut-off should be considered in the grey zone.
   - It is recommended to repeat test again 2-4 weeks later with fresh sample. If the results in the second test are again in the grey zone, the sample has to be considered negative.
   - Samples are considered negative if the absorbance value is lower than 10% below the cut-off.

**QUALITY ASSURANCE**

*Intra-assay and Inter-assay precision*

The CV value of intra-assay precision is 3.1% and inter-assay precision is 4.8%.
**Specificity**

No cross reactivity was observed with the following factors:

- Adenovirus, CMV, EBV, HBV, Echinococcus, Influenza, Leptospira,
- Mycoplasma, Picorna, RSV, Rubella, Syphilis, Toxoplasma, VZV
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