

IL-6 ELISA Cat. No. BI-IL6 12x8 Tests

IMMUNOASSAY FOR THE QUANTITATIVE DETERMINATION OF HUMAN INTERLEUKIN-6 (IL-6) IN SERUM, PLASMA, CELL CULTURE SUPERNATANTS, AND URINE

For research use only. Not for use in diagnostic procedures.

CONTENTS

ASSAY CHARACTERISTICS OVERVIEW	2
ASSAY PRINCIPLE	3
TYPICAL DATA	4
CALIBRATION	4
DETECTION LIMIT & SENSITIVITY	5
PRECISION	5
Within-Run Precision	5
In-Between-Run Precision	5
ACCURACY	6
DILUTION LINEARITY & PARALLELISM	7
Parallelism	7
Dilution Linearity	9
SPECIFICITY	10
Competition of Signal	10
CROSS REACTIVITY with non-human samples	11
SAMPLE STABILITY	11
Freeze-thaw Stability	11
Benchtop Stability	12
SAMPLE VALUES	13
IL-6 Values in Apparently Healthy Individuals	13
IL-6 Values in Disease Panels	
MATRIX COMPARISON	14
Measurement of Human IL-6 in Urine Samples	15
Measurment of Human IL-6 in Cell Culture Supernatants (CCS)	
REFERENCES & DOCUMENTS	17
COMPARISON with another human II -6 FLISA assay	18



ASSAY CHARACTERISTICS OVERVIEW

Method	Sandwich ELISA, HRPO/TMB, 12x8-well detachable strips								
Sample type(s)	Serum, plasma (EDTA, citrate, heparin), cell culture supernatants, urine								
Sample volume	100 μl sample / well								
Standard range	0 - 200 pg/ml (0 / 3.12 / 6.25 / 12.5 / 25 / 50 / 100 / 200)								
Sensitivity		LOD: 0.28 pg/ml; LLOQ: 0.78 pg/ml (measurable concentrations in serum AND plasma samples!)							
Assay time	2h/1h/1h/30 mi	n							
Conversion	1 pg/ml=0.048 pmo	I/I (MW=20.8	kD)						
Duration		n			Average	e % CV			
Precision	Within-run	3			≤ 7	7			
	In-between-run				In prog	gress			
		n		J	Average %	recovery			
				+100 pg/ml		+50 pg/ml			
Accuracy	Serum	6	6		13	112			
(Spike/Recovery of	EDTA plasma	6		111		109			
recombinant human IL-6)	Citrate plasma	2		111		99			
	Heparin plasma	2		1	07	103			
	Cell culture	3		n.d.		97			
	Urine	5		n.d.		102			
			Average % of		% of expe	cted dilution			
		n		1+1	1+3	1+7			
Parallelism of	Serum	5		93	90	89			
endogenous human	EDTA plasma	5		99	96	92			
IL-6	Citrate plasma	2		103	100	99			
IL-0	Heparin plasma	2		104	90	89			
	Cell culture	2		95	102	98			
	Urine	2		96	108	108			
Specificity	This assay recognize	es recombina	nt and	d endoge	nous (natu	ıral) human IL-6.			
Use	Research use only.								
	,	n		Med	dian / Rang	ge IL-6 pg/ml			
Values of	Serum	48			1.50 (0.3				
Values of	EDTA plasma	26			0.98 (0.0	1-2.69)			
apparently healthy	Citrate plasma	14			0.71 (0.0	1-2.10)			
donors	Heparin plasma	11			0.60 (0				
				0.80 (0-2.41)					
	Urine	4			0.77 (0	-1.5)			

Abbreviation: n.d.: not determined

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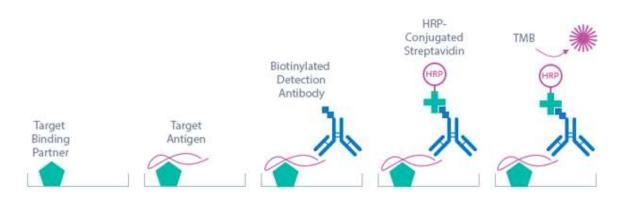




ASSAY PRINCIPLE

The Biomedica human Interleukin-6 ELISA (IL-6) ELISA kit is a sandwich enzyme immunoassay that has been optimized and fully validated for the quantitative determination of human IL-6 in serum and plasma (EDTA, heparin, citrate). Validation experiments have been performed according to international quality guidelines (ICH/ FDA/ EMEA). Cell culture supernatant and urine samples are compatible with this ELISA (data download: www.bmgrp.com). The IL-6 ELISA assay recognizes both natural and recombinant human IL-6. The assay employs highly purified epitope mapped antibodies as well as human serum-based standards and controls.

The figure below explains the principle of the human IL-6 sandwich ELISA:

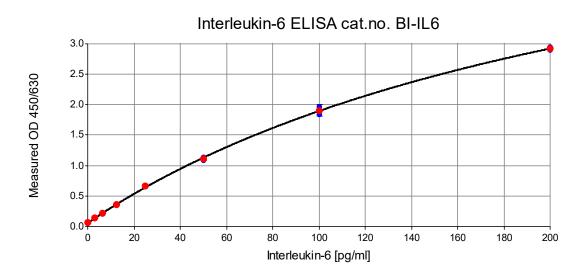


In a first step, STD/sample/CTRL are pipetted into the wells, which are pre-coated with the recombinant anti-human IL-6 antibody. Any soluble IL-6 present in the STD/sample/CTRL binds to the pre-coated anti-IL-6 antibody in the well. After incubation, a washing step is applied where all non-specific unbound material is removed. In a next step, the biotinylated anti-IL-6 antibody (AB) is pipetted into the wells and reacts with the IL-6 present in the sample, forming a sandwich. Next, all unbound antibody is removed during another washing step. In the next step, the conjugate (streptavidin-HRPO) is added and reacts with the biotinylated anti-IL-6 antibody. After another washing step, the substrate (Tetramethylbenzidine; TMB) is pipetted into the wells. The enzyme catalysed color change of the substrate is directly proportional to the amount of IL-6 present in the sample. This color change is detectable with a standard microtiter plate ELISA reader. A dose response curve of the absorbance (optical density, OD at 450 nm) versus standard concentration is generated, using the values obtained from the standards. The concentration of soluble IL-6 in the sample is determined directly from the dose response curve.



TYPICAL DATA

This standard curve and the displayed OD values are for demonstration only. A standard curve should be generated for each assay run.



Standard	IL-6 [pg/ml]			CV [%]	
otaniaan a	.= 0 [68,]			Average	0. [50]
STD1	0	0.056	0.059	0.058	4
STD2	3.125	0.140	0.129	0.135	6
STD3	6.25	0.216	0.207	0.212	3
STD4	12.5	0.354	0.366	0.360	2
STD5	25	0.661	0.660	0.661	0
STD6	50	1.090	1.138	1.114	3
STD7	100	1.970	1.835	1.903	5
STD8	200	2.951	2.894	2.923	1

The quality control protocol supplied with the kit shows the results of the final release QC for each kit at the production date. ODs obtained by customers may differ due to various influences including a normal decrease of signal intensity throughout shelf life. However, this does not affect the validity of the results, provided an OD of 1.50 or higher is obtained for the standard with the highest concentration, and the measured control values fall into their respective target range (see labels).

CALIBRATION

The Biomedica human Interleukin-6 (IL-6) ELISA kit is calibrated against a highly purified recombinant human IL-6 protein (expressed in human embryonic kidney cell, HEK-293). The human serum based calibrator is provided in eight lyophilized glass vials in the following concentrations: 0/3.125/6.25/12.5/25/50/100/200 pg/ml.

CALIBRATION using WHO standard

The WHO reference reagent IL-6/NIBSC code 89/548 (recombinant DNA, human sequence) was analysed in this human IL-6 ELISA kit.



The equation below can be used to convert the sample values obtained with this kit to approximate WHO/IL-6 /NIBSC 89/548 units:

WHO/NIBSC (89/548) reference (IU/mI) = 0.08 BI-IL6 value (pg/mI).

DETECTION LIMIT & SENSITIVITY

To determine the sensitivity of the IL-6 ELISA, experiments measuring the Lower Limit of Detection (LOD) and the Lower Limit of Quantification (LLOQ) were conducted.

The LOD, also called the detection limit, is the lowest point at which a signal can be distinguished from the background signal, i.e., the signal that is measured in the absence of IL-6, with a confidence level of 99%. It is defined as the mean back-calculated concentration of standard 1 (0 pg/ml of IL-6, five independent measurements) plus three times the standard deviation of the measurements.

The LLOQ, or sensitivity of an assay, is the lowest concentration at which an analyte can be accurately quantified. The criteria for accurate quantification at the LLOQ are an analyte recovery between 75% and 125% and a coefficient of variation (CV) of less than 25%. To determine the LLOQ, standard 2, *i.e.*, the lowest standard containing IL-6, is diluted, measured five times and its concentration back calculated. The lowest dilution, which meets both criteria, is reported as the LLOQ.

The following values were determined for the human IL-6 ELISA:

LOD	0.28 pg/ml
LLOQ	0.78 pg/ml

PRECISION

The precision of an ELISA is defined as its ability to measure the same concentration consistently within the same experiments carried out by one operator (within-run precision or repeatability) and across several experiments using the same samples but conducted by several operators at different locations using different ELISA lots (in-between-run precision or reproducibility).

Within-Run Precision

Within-run precision was tested by measuring two samples of known concentrations three times within one IL-6 ELISA lot by one operator.

ID	n	Mean IL-6 [pg/ml]	SD IL-6 [pg/ml]	CV [%]
Sample 1	3	6.5	0.5	7
Sample 2	3	50.5	0.7	1

In-Between-Run Precision

In progress



ACCURACY

The accuracy of an ELISA is defined as the precision with which it can recover samples of known concentrations.

The recovery of the human Interleukin-6 (IL-6) ELISA was measured by adding recombinant human IL-6 to samples containing a known concentration of endogenous IL-6. The %recovery of the spiked concentration was calculated as the percentage of measured over the expected value.

This table shows the summary of the recovery experiments in the IL-6 ELISA in different sample matrices:

		% Recovery [recombinant IL-6]]						
Sample Matrix	_	+100 pg/ml		+:	50 pg/ml	+25 pg	+25 pg/ml	
	n	Mean	Range	Mean	Range	Mean	Range	
Serum	6	113	103-122	112	103-120	96	88-103	
EDTA plasma	6	111	107-116	109	103-113	99	93-104	
Citrate plasma	2	111	106-115	99	97-102	102	99-105	
Heparin plasma	2	107	105-109	103	103-104	95	89-100	
Cell culture	2	n.d.	n.d.	97	90-103	_	_	
supernatant		n.u.	n.u.	//	70 103			
Urine	5	n.d.	n.d.	102	95-110	-	-	

n.d.: not determined

Experiments:

Recovery of spiked samples was tested by adding various concentrations of human recombinant IL-6 to different human sample matrices.

Data showing % recovery of recombinant IL-6 in human serum samples:

ID		IL-	-6		% Recovery			
ID	Reference	+100 pg/ml	+50 pg/ml	+25 pg/ml	+100 pg/ml	+50 pg/ml	+25 pg/ml	
S1	2.7	104.4	52.8	26.6	103	103	97	
S2	2.5	114.1	53.7	24.8	113	105	90	
S3	1.4	119.7	57.6	26.1	119	114	99	
S4	2.5	113.7	61.1	26.4	112	120	97	
S5	0.5	122.3	56.9	26.2	122	113	103	
S6	17.7	116.2	66.8	37.6	107	116	88	
				Mean R[%]	113	112	96	

Data showing % recovery of recombinant IL-6 in human EDTA plasma samples:

ID		IL-	-6	% Recovery			
ID Reference	Reference	+100 pg/ml	+50 pg/ml	+25 pg/ml	+100 pg/ml	+50 pg/ml	+25 pg/ml
E1	2.1	110.7	57.4	26.1	110	113	97
E2	2.7	116.9	52.8	25.5	116	103	93
E3	0.7	107.6	55.5	25.3	107	110	99
E4	2.9	113.4	56.0	27.2	112	109	99
E5	0.6	111.1	55.2	26.4	111	110	104
E6	0.3	110.1	55.4	25.5	110	111	101
				Mean R[%]	111	109	99



Data showing % recovery of recombinant IL-6 in human citrate plasma samples:

		IL-6				% Recovery			
ID	Reference +100 pg/ml +50 pg/ml		+25 pg/ml	+100 pg/ml	+50 pg/ml	+25 pg/ml			
C1	2.1	115.6	52.2	28.1	115	102	105		
C2	1.7	107.3	49.1	26.3	106	97	99		
•				Mean R[%]	111	99	102		

Data showing % recovery of recombinant IL-6 in human heparin plasma samples:

ID		IL-6				% Recovery			
טו	Reference	+100 pg/ml	+50 pg/ml	+25 pg/ml	+100 pg/ml	+50 pg/ml	+25 pg/ml		
H1	0.9	105.8	52.2	25.8	105	104	100		
H2	0.9	109.4	51.7	23.1	109	103	89		
				Mean R[%]	107	103	95		

DILUTION LINEARITY & PARALLELISM

Tests of dilution linearity and parallelism ensure that both, endogenous and recombinant samples containing IL-6 behave in a dose-dependent manner and are not affected by matrix effects. Dilution linearity assesses the accuracy of measurements in diluted human samples spiked with known concentrations of recombinant analyte. By contrast, parallelism refers to dilution linearity in clinical samples and provides evidence that endogenous analyte behaves in the same way as the recombinant one /likewise to the recombinant analyte. Dilution linearity and parallelism are assessed for each sample type and should be within 20% of the expected concentration.

Parallelism

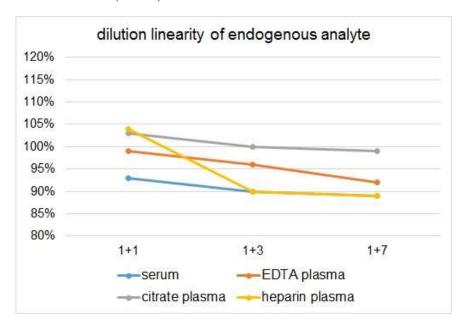
Experiment:

Parallelism was assessed by serially diluting human samples containing **endogenous** IL-6 with assay buffer.

Summary table below shows the mean recovery and range of serially diluted endogenous IL-6 in several sample matrices:

		% Recovery of endogenous IL-6 in diluted samples							
Comple Matrix	_	1+1		1	.+3	1+7			
Sample Matrix	n	Mean	Range	Mean	Range	Mean	Range		
Serum	5	93	82-100	90	86-94	89	84-93		
EDTA plasma	5	99	92-104	96	76-114	92	86-98		
Citrate plasma	2	103	101-106	100	92-109	99	92-106		
Heparin plasma	2	104	104-104	90	84-95	89	88-90		
Cell culture supernatant	2	95	89-101	102	88-117	98	88-108		
Urine	2	96	95-98	108	97-120	108	93-124		





Data showing dilution linearity of endogenous IL-6 in human serum samples:

ID		IL-6 [pg	% Recovery				
ן טו	Reference	1+1	1+3	1+7	1+1	1+3	1+7
S1	40.3	19.4	9.1	4.7	96	90	93
S2	44.4	18.1	9.9	4.9	82	89	88
S3	91.9	41.6	21.5	10.0	91	94	87
S4	39.9	20.0	9.1	4.2	100	91	84
S5	45.8	22.3	9.6	5.2	97	86	91
				Mean R[%]	93	90	89

Data showing dilution linearity of endogenous IL-6 in human EDTA plasma samples:

ID		IL-6 [pg	% Recovery				
טו	Reference	1+1	1+3	1+7	1+1	1+3	1+7
E1	74.2	34.1	14.1	8.2	92	76	88
E2	22.0	11.0	5.2	2.7	100	95	98
E3	38.1	18.1	8.8	4.1	95	92	86
E4	9.8	5.0	2.8	1.1	102	114	90
E5	31.4	16.3	8.0	3.8	104	102	97
		Mean R[%]	99	96	92		

Data showing recovery of endogenous IL-6 in human citrate plasma samples:

ID		IL-6 [pg	% Recovery				
ID	Reference	1+1	1+3	1+7	1+1	1+3	1+7
C1	59.3	29.9	13.7	6.8	101	92	92
C2	147.8	78.2	40.1	19.5	106	109	106
		Mean R[%]	103	100	99		

Data showing recovery of endogenous IL-6 in human heparin plasma samples:

ID		IL-6 [p		% Recovery			
ID	Reference	1+1	1+3	1+7	1+1	1+3	1+7
H1	144.7	75.1	34.5	16.0	104	95	88
H2	92.1	48.0	19.4	10.4	104	84	90
				Mean R[%]	104	90	89



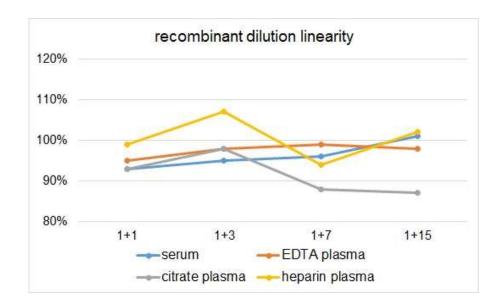
Dilution Linearity

Experiment:

Dilution linearity was assessed by serially diluting samples containing **recombinant** human IL-6 with assay buffer.

The figure and table below show the mean recovery and range of serially diluted recombinant IL-6 in several sample matrices:

		% Recovery of recombinant IL-6 in diluted samples								
Commis Matrix	_	1+1		1	l+3	1+7				
Sample Matrix	n	Mean	Range	Mean	Range	Mean	Range			
Serum	5	93	83-100	95	85-104	96	91-104			
EDTA plasma	5	95	85-104	98	93-104	99	94-106			
Citrate plasma	2	93	93-94	98	94-102	88	86-89			
Heparin plasma	2	99	94-104	107	103-111	94	94-94			
Cell culture supern	2	93	90-97	95	92-97	96	94-98			
Urine	1	89	-	99	-	89	-			



Data showing dilution linearity of recombinant IL-6 spiked into human serum and plasma samples (reference) containing endogenous IL-6.

Calculation of dilution linearity of spiked serum samples:

ID		IL-6 [pg	% Recovery				
טו	Reference	1+1	1+3	1+7	1+1	1+3	1+7
S1	126.8	63.1	30.9	14.7	100	97	93
S2	114.1	50.6	26.3	14.8	89	92	104
S3	119.7	57.7	29.0	14.3	96	97	96
S4	113.7	54.7	29.7	14.3	96	104	96
S5	124.7	51.6	26.4	14.2	83	85	91
				Mean R[%]	93	95	96



Calculation of dilution linearity of spiked EDTA plasma samples:

ID		IL-6 [pg	% Recovery				
טו	Reference	1+1	1+3	1+7	1+1	1+3	1+7
E1	110.7	57.8	28.9	14.7	104	104	106
E2	116.9	49.5	28.3	14.2	85	97	97
E3	111.1	50.4	25.8	13.4	91	93	96
E4	111.4	52.7	26.0	14.1	95	93	101
E5	119.3	60.7	30.7	14.0	102	103	94
		•	•	Mean R[%]	95	98	99

Calculation of dilution linearity of spiked citrate plasma samples:

ID		% Recovery					
ID	Reference	1+1	1+7	1+1	1+3	1+7	
C1	115.6	54.2	29.4	12.9	94	102	89
C2	107.3	49.9	25.2	11.5	93	94	86
		Mean R[%]	93	98	88		

Calculation of dilution linearity of spiked heparin plasma samples:

ID		IL-6 [pg	% Recovery				
טו	Reference	1+1	1+3	1+7	1+1	1+3	1+7
H1	105.8	49.6	27.2	12.4	94	103	94
H2	109.4	56.8	30.3	12.9	104	111	94
				Mean R[%]	99	107	94

SPECIFICITY

The specificity of an ELISA is defined as its ability to exclusively recognize the analyte of interest.

The specificity of the IL-6 ELISA was shown by characterizing both the capture and the detection antibodies through epitope mapping. In addition, the specificity of the ELISA was established through competition experiments, which measure the ability of the antibodies to exclusively bind IL-6.

This assay recognizes recombinant and endogenous (natural) human IL-6 and detects free circulating IL-6 as well as receptor-bound IL-6.

The recombinant anti-human IL-6 capture antibody detects a structural epitope near the receptor binding site of the IL-6 molecule. Four linear epitopes of the polyclonal anti-human IL-6 detection antibody are spread throughout the IL-6 protein.

Competition of Signal

Competition experiments were carried out by pre-incubating human samples containing endogenous IL-6 with an excess of capture antibody (AB). The concentration measured in this mixture was then compared to a reference value, which was obtained from the same sample without the pre-incubation step. Mean competition in serum and plasma samples is 99%.



		IL-e	IL-6 [pg/ml]		
Sample Matrix	ID	Reference	+ capture AB	% Competition	
Serum	S1	1.4	0.0	95	
Serum	S2	61.4	0.3	99	
Serum	S 3	41.5	0.3	99	
Serum	S4	33.2	0.6	98	
EDTA plasma	E1	18.9	0.2	99	
EDTA plasma	E2	27.6	0.1	99	
Citrate plasma	C1	45.6	0.0	100	
Heparin plasma	H1	81.0 0.0		100	
	•	_	Mean R[%]	99	

CROSS REACTIVITY with non-human samples

This human IL-6 ELISA kit cannot be used for the detection of IL-6 in rat, mouse, and porcine samples.

SAMPLE STABILITY

Serum, EDTA plasma, heparin plasma, citrate plasma, cell culture supernatants, and urine samples are suitable for use in this assay. Do not change sample type during studies. We recommend duplicate measurements for all samples, standards and controls. The sample collection and storage conditions listed are intended as general guidelines.

Freeze-thaw Stability

The stability of endogenous Interleukin-6 (IL-6) was tested by comparing samples that had undergone five freeze-thaw cycles (F/T).

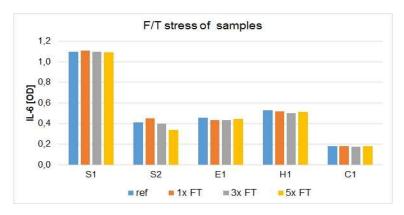
For freeze-thaw experiments, samples were collected according to the supplier's instruction using blood collection devices and stored at -80°C. Reference samples were freeze-thawed once. The mean recovery of sample concentration after five freeze-thaw cycles is 95%.

Sample OD values after freeze-thaw (F/T) cycles:

			IL-6	[OD]		% Recovery			
Sample Matrix	ID	Reference	1x F/T	3x F/T	5x F/T	1x F/T	3x F/T	5x F/T	
Serum	S1	1.09	1.10	1.09	1.09	101	100	100	
Serum	S2	0.41	0.45	0.43	0.33	110	98	82	
EDTA plasma	E1	0.45	0.43	0.43	0.44	95	95	98	
Citrate plasma	C1	0.18	0.18	0.17	0.18	98	95	99	
Heparin plasma	H1	0.53	0.52	0.50	0.51	98	94	97	
				·	Mean R[%]	101	97	95	

All samples should undergo a maximum of five freeze-thaw cycles.





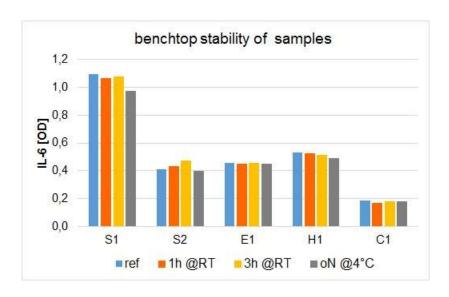
Benchtop Stability

The benchtop stability of endogenous Interleukin-6 (IL-6) was tested by comparing IL-6 measurements in human samples that had been stored at different temperatures.

For the assessment of the benchtop stability, a set of human samples was aliquoted and stored at room temperature or at 4°C. Samples can be stored for at least three hours at room temperature as well as overnight at 4°C. The mean recovery of sample concentrations after overnight storage at 4°C is 95%.

Sample OD values of samples stored at -25° C (reference), at room temperature (RT) or overnight (o.n.) at 4° C:

			IL-6 (OD)		% Recovery		
Sample Matrix	ID	Reference	3h @RT	o.n. @4°C	3h@RT	o.n. @4°C	
Serum	S1	1.09	1.07	0.97	98	89	
Serum	S2	0.41	0.46	0.39	114	97	
EDTA plasma	E1	0.45	0.45	0.44	99	98	
Citrate plasma	C1	0.18	0.17	0.17	96	97	
Heparin plasma	H1	0.53	0.51	0.49	96	92	
				Mean R [%]	101	95	





SAMPLE VALUES

IL-6 Values in Apparently Healthy Individuals

To provide values for circulating Interleukin-6 (IL-6), a panel of samples from apparently healthy donors was tested (no medical histories were available). Each individual donated blood for all tested sample matrices.

			% Detectable		
Sample Matrix	n	Mean	Range	Median	% Detectable
Serum	48	1.73	0.30 - 4.36	1.50	100
EDTA plasma	26	1.01	0.01 - 2.69	0.98	100
Citrate plasma	14	1.86	0.01 - 2.10	0.71	100
Heparin plasma	11	1.52	0.00 - 2.41	0.60	91

It is recommended to establish the normal range for each laboratory.

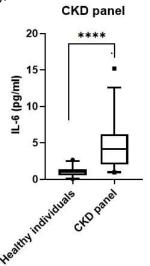
IL-6 values measured in serum and plasma (EDTA, citrate, heparin) samples of ten apparently healthy donors (matched-paired samples):

			% Detectable		
Sample Matrix	n	Mean	Range	Median	70 Detectable
Serum	10	1.12	0.58-1.56	1.19	100
EDTA plasma	10	0.64	0.01-1.45	0.53	100
Citrate plasma	10	0.71	0.01-1.32	0.68	100
Heparin plasma	9	0.62	0.00-2.41	0.48	90

IL-6 Values in Disease Panels

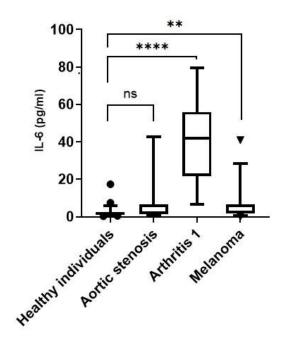
In addition to samples of apparently healthy donors, panels of samples from patients with heart disease (aortic stenosis), kidney diseases, as well as a panel of unselected hospital patients were tested.

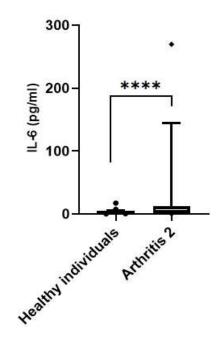
IL-6 values measured in apparently healthy individuals and in subjects with chronic kidney disease (CKD).





IL-6 values measured in apparently healthy individuals and in subjects with aortic stenosis, arthritis, and melanoma.





Summary of the results:

,		IL-6 [pg/ml]			
Samples / Matrix	n	Mean	Range	Median	
Apparently Healthy Panel / Serum	48	1.7	0.3-4.3	1.5	
Aortic Stenosis Panel / Serum	18	6.5	0.8-42.6	2.0	
Arthritis Panel I / Serum	13	40.5	6.5-79.4	41.9	
Arthritis Panel II / Serum	32	18.5	0.4-270	3.4	
Melanoma Panel / Serum	32	6.0	0-40.9	2.5	
Nephro-CKD Panel / EDTA plasma	28	4.5	1.0-15.2	4.1	

MATRIX COMPARISON

To assess whether all tested matrices behave the same way Interleukin-6 (IL-6) was measured in serum and plasma (EDTA, citrate, heparin) samples of ten apparently healthy donors (matched-paired samples).

			% Detectable		
Sample Matrix	n	Mean	Range Median		% Detectable
Serum	10	1.12	0.58-1.56	1.19	100
EDTA plasma	10	0.64	0.01-1.45	0.53	100
Citrate plasma	10	0.71	0.01-1.32	0.68	100
Heparin plasma	10	0.62	0.00-2.41	0.48	90



Measurement of Human IL-6 in Urine Samples

Aseptically collect the first urine of the day (mid-stream), voided directly into a sterile container. Centrifuge to remove particles, assay immediately or aliquot and store at -25°C or lower. Urine samples were not normalized to creatinine values.

Sample Matrix Urine	IL-6 [pg/ml]
Donor 1. Apparently healthy	0.9
Donor 2. Apparently healthy	0.7
Donor 3. Apparently healthy	1.5
Donor 4. Apparently healthy	0.0
Donor 1. Kidney disease	122.6
Donor 2. Kidney disease	24.6
Donor 3. Kidney disease	36.6
Donor 4. Kidney disease	2.1
Donor 5. Kidney disease	13.9

Acccuracy

Data showing % recovery of recombinant IL-6 in human urine samples:

ID	IL-6 [p	9/ Deceytory	
טו	Reference	+50 pg/ml	% Recovery
U1	13.9	60.5	107
U2	41.7	80.7	120
U3	0.7	60.7	121
U4	1.5	58.3	115
U5	0.0	59.4	119
	_	Mean R [%]	116

Dilution linearity, parallelism

Data showing dilution linearity of endogenous IL-6 in human urine samples:

		IL-6 [pg/ml]	R[%]	
Sample ID	Ref	1+1	1+3	1+1	1+3
U1	124.8	54.8	27.0	88	86
U2	13.9	7.2	2.6	103	n.d.
U3	41.7	24.9	12.0	120	115
			Mean R [%]	103	101

Competition experiments were carried out by pre-incubating human urine samples containing endogenous IL-6 with an excess of capture antibody (AB). The concentration measured in this mixture was then compared to a reference value, which was obtained from the same sample without the pre-incubation step. Mean competition in urine samples was 100%.



Competition of endogenous signal:

Sample ID	IL-6 [pg/	R [%]		
Sample ID	Reference	+AB	K [%]	
U1	147	0.0	100	
U2	16.6	0.0	100	
U3	28.6	0.0	100	
U4	36.1	0.0	100	
U5	9.5	0.0	100	
		Mean R [%]	100	

Measurment of Human IL-6 in Cell Culture Supernatants (CCS)

Two human breast cancer cell lines MDA-MB-231, MCF-7 and a human macrophage cell line 4TL9.R were cultured in DMEM/Ham's F12 and RPMI, respectively, and supplemented with 10% fetal bovine serum and 1% penicillin/streptomycin. Cells were grown in a humidified atmosphere of 95% air and 5% CO₂ for 48 hours. Aliquots of the cell culture supernatants were removed, centrifuged to remove particles, and assayed for levels of human IL-6.

Sample Matrix CCS	IL-6 [pg/ml]
CCS - MDA-MB-231	146.6
CCS - MCF-7	18.5
CCS - 4TL9.R	0.4
Medium- DM-F-12 (with supplements)	0.0
Medium-RPMI (with supplements)	0.0

Accuracy

Recombinant IL -6 was spiked into samples by using STD7 (100pg/ml) or STD8 (200pg/ml).

Two concentration levels were generated using the following protocol:

-50 pg/ml spike: STD7 volume ratio of 1+1.

-25 pg/ml spike: STD8 volume ratio of 7+1.

ccs	ID	IL-	R [%]			
	.5	Reference	50	25	50	25
MDA-MB-231	CCS#1	146.6	118.2	n.d.	90	n.d.
MCF-7	CCS#2	18.5	57.9	40.7	97	98

n.d.: not determined

Dilution linearity

Dilution linearity of recombinant analyte in cell culture supernatants. Samples were spiked with STD 7 (100pg/ml, +50pg/ml) and diluted with dilution medium (ASYBUF).

ccs		[pg/ml]	% Recovery					
CCS	ID	Ref	1+1	1+3	1+7	1+1	1+3	1+7
MDA-MB-231	CCS#1	118.2	53.1	27.1	13.9	90	92	94
MCF-7	CCS#2	57.9	28.0	14.1	7.1	97	97	98
					Mean R [%]	93	95	96



Dilution linearity of endogenous analyte in cell culture supernatants.

Performance was tested in conditioned media (48h) and diluted with dilution medium (ASYBUF).

ccs	ID	IL-6 [pg/ml]			% Recovery			
CCS	שו	Ref	1+1	1+3	1+7	1+1	1+3	1+7
MDA-MB-231	CCS#1	146.6	65.0	32.2	16.1	89	88	88
MCF-7	CCS#2	18.5	9.3	5.4	2.5	101	117	108
					Mean R [%]	95	102	98

Competition of IL-6 in cell culture supernatants

Competition experiments were carried out by pre-incubating human cell culture supernatnant samples containing endogenous IL-6 with an excess of capture antibody (AB). The concentration measured in this mixture was then compared to a reference value, which was obtained from the same sample without the pre-incubation step. Mean competition in cell culture supernatants was 98%.

Calculated according to calibration curve prepared from supplied ELISA standards (resuspended in DMEM Medium + supplements).

ccs	ID		IL-6	R [%]
CC3		Ref	+ CAB	K [70]
MDA-MB-231	CCS#1	146.6	0.4	100
MCF-7	CCS#2	18.5	0.8	96
			Mean R [%]	98

REFERENCES & DOCUMENTS

Validation Literature

The assay is fully validated according to:

- 1. ICH Topic Q2 (R1) "Validation of Analytical Procedures: Text and Methodology"
- 2. EMEA/CHMP/EWP/192217/2009 Guideline on bioanalytical method validation
- 3. Bioanalytical Method Validation, Guidance for Industry, FDA, May 2018

Additional Documents Available Online (<u>www.bmgrp.com</u>)

Instructions for Use (IFU, package insert)
Material Safety Data Sheet (MSDS)

This ELISA kit was developed and manufactured by:

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COMPARISON with another human IL-6 ELISA assay

	BIOMEDICA	Another MANUFACTURER	
Method	Sandwich ELISA Streptavidin-HRPO/TMB, 12x8-well detachable strips	Sandwich ELISA HRPO/TMB, 12x8-well detachable strips	
Sample type	Serum, EDTA plasma, heparin plasma, citrate plasma, cell culture supernatants, urine	Serum, plasma, cell culture supernates	
Sample volume	100 μl sample / well	100 µl / well 200 µl (cell culture) / well	
Assay time	2 h / 1 h / 1 h / 30 min	2 h / 2 h / 30 min	
Assay range	0 -200 pg/ml (0 / 3.125 / 6.25 / 12.5 / 25 / 50 / 100 / 200)	0 – 300 pg/ml	
Sensitivity	LOD: 0.28 pg/ml; LLOQ: 0.78 pg/ml (measurable concentrations in serum AND plasma samples)	MDD: 0.7 pg/ml	
Specificity	Assay recognizes recombinant and endogenous (natural) human IL-6.	Assay recognizes natural and recombinant human IL-6.	
Antibodies	Epitope-mapped antibodies Capture antibody: recombinant IL-6 antibody specific for human IL-6 Detection antibody: polyclonal IL-6 antibody specific for human IL-6, streptavidin-HRPO-labeled	Capture antibody: monoclonal antibody specific for human IL-6 Detection antibody: polyclonal IL-6 antibody specific for human IL-6, HRPO-labeled	
Standard matrix	Serum based matrix containing recombinant IL-6 8 ready to use standards, lyophilized	Protein based matrix containing recombinant IL-6 1 stock standard, lyophilized	
Values of apparently healthy samples	Serum median (n=48): 1.5 pg/ml 100 % detectable EDTA-plasma mean (n=26): 0.9 pg/ml 100 % detectable	Serum/plasma median (n=40): 33 samples < 3.13 pg/ml 3.13 pg/ml < 7 samples < 12.5 pg/ml	
Controls	2 controls (high and low) included	Not included	
Validation	According to FDA/ICH/EMEA guidelines	Not indicated	
Use	RUO	RUO	



Comparison of human sample concentrations measured with different human IL-6 ELISA Assays

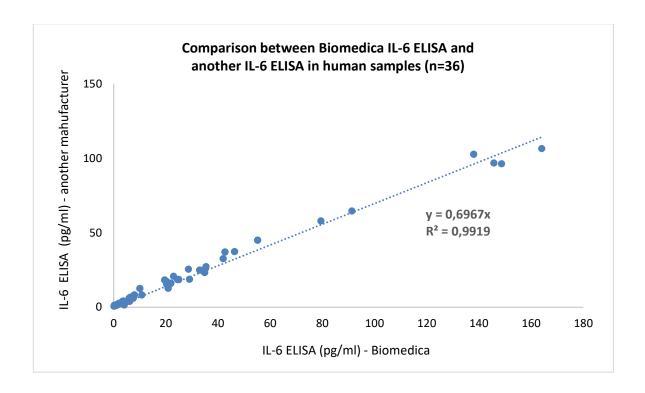
The Biomedica human IL-6 ELISA kit (Cat. No. BI-IL6) was compared with an ELISA kit from another manufacturer. The same panel of samples, consisting of 36 samples (healthy and diseased), were tested.

Table showing human IL-6 concentrations measured with the Biomedica human IL-6 ELISA and a human IL-6 ELISA assay from another manufacturer:

	ALL SAMPLES		
	n = 36		
Cohorts:	Sample ID	Biomedica (BI-IL6)	Other
		IL-6 [pg/ml]	IL-6 [pg/ml]
Apparently	AHs1	0.1	0.6
healthy cohort	AHs2	1.1	1.3
	AHs3	3.5	3.3
serum samples	AHs4	0.2	1.3
	AH s5	1.0	1.3
	AH s6	2.4	2.6
	AH s7	0.5	0.9
	AH s8	1.2	1.8
	AHs9	1.9	2.5
	Mean	1.31	1.75
Rheuma cohort	Rs1	46.3	37.3
	Rs2	41.9	32.6
serum samples	Rs3	55.1	45.0
	Rs4	22.9	20.7
	Rs5	20.8	12.7
	Rs6	79.4	58.0
	Mean	44.4	34.4
Cardiology panel	Cs1	3.4	4.1
(Aortic stensosis)	Cs2	6.4	6.7
serum samples	Cs3	7.9	8.2
	Cs4	42.6	37.0
	Cs5	28.6	25.5
	Mean	17.8	16.3
Unselected	Us1	24.4	18.5
hospital	Us2	29.0	18.7
panel	Us3	24.9	18.5
	Us4	6.1	3.9
Serum samples	Us5	34.8	23.3
	Us6	21.9	16.1



Unselected	Mean	23.5	16.5
hospital	Uep1	19.5	18.2
panel	Uep2	10.0	12.6
	Uep3	7.5	6.1
EDTA plasma	Uep4	4.9	3.6
	Uep5	34.2	24.4
	Uep6	34.7	25.1
	Mean	18.5	15.0
Unselected	Uc1	91.3	64.6
hospital	Uc2	20.3	15.4
panel	Uc3	145.7	96.8
	Uc4	10.7	8.3
Citrate plasma	Mean	67.0	46.3



IL-6 ELISAs Comparison - Conversion to NIBSC/WHO units

Concentrations of samples obtained in the Biomedica IL-6 ELISA and the competitor IL-6 ELISA were converted to approximate NIBSC/WHO 89/548 units according to the equation:

- NIBSC/WHO 89/548 approximate value (IU/ml) = $0.131 \, x$ obtained value (pg/ml) in competitor II -6 FLISA
- NIBSC/WHO 89/548 approximate value (IU/ml) = 0.08 x obtained value (pg/ml) in Biomedica IL-6 ELISA



	Biomedica IL-6 ELISA (#BI-IL6)	Competitor IL-6 ELISA	ratio
Sample ID	IL-6 [pg/ml]	IL-6 [pg/ml]	
#S1	1.95	2.42	0.81
#S2	2.32	2.45	0.95
#S3	1.99	2.42	0.82
#S4	0.49	0.51	0.95
#S5	2.79	3.05	0.91
#S6	1.75	2.11	0.83
#S7	3.70	4.89	0.76
#S8	3.35	4.27	0.79
#S9	4.41	5.89	0.75
#S10	1.83	2.71	0.68
#S11	1.66	1.67	1.00
#S12	6.35	7.59	0.74
#S13	0.27	0.53	0.51
#S14	0.51	0.88	0.58
#S15	0.63	1.07	0.59
#S16	3.41	4.85	0.70
#S17	2.29	3.34	0.69
#H1	2.83	3.57	0.79
#H2	7.45	14.08	0.53
#H3	0.47	0.8	0.59
#E1	1.56	2.39	0.65
#E2	0.80	1.65	0.48
#E3	0.60	0.80	0.75
#E4	0.39	0.48	0.82
#E5	2.74	3.19	0.86
#E6	2.78	3.29	0.84
#E7	11.03	13.45	0.82
#C1	7.30	8.46	0.86
#C2	1.62	2.02	0.80
#C3	11.65	12.69	0.92
#C4	0.86	1.08	0.79
#U1	9.81	12.2	0.80
#U2	1.97	2.99	0.66
#U3	0.32	0.52	0.61
#CCM1	13.12	13.95	0.94
#CCM2	1.63	2.26	0.72

Abbreviations: S (serum), H (heparin plasma), E (EDTA plasma), C (citrate plasma), U (urine), CCM (cell culture supernatants)

Conclusion:

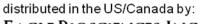
Concentration values obtained in the Biomedica IL-6 ELISA converted to IU/ml using NIBSC/WHO 89/548 measure very similar to the competitor IL-6 ELISA in all matrices. Correlation of both assays is excellent – Pearson correlation coefficient R = 0.978, p < 0.00001.



Comparison of cell culture supernatant sample concentrations in human breast cancer cell lines measured with different human IL-6 ELISA Assays

The Biomedica IL-6 ELISA (#BI-IL6) was compared with an ELISA kit from another manufacturer.

Cell Culture Supernatants (CCS)			
n = 2			
Sample ID	Biomedica (# BI-IL6)	Other ELISA	
	IL-6 [pg/ml]	IL-6 [pg/ml]	
CCS - MDA-231	164.0	106.5	
CCS - MCF-7	20.3	17.3	



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